

Índex dels annexos

Índex dels annexos	1
Índex de figures	3
Índex de taules	5
A. <i>Electrospinning</i> de dissolucions i polímers fosos	7
B. Caracterització dels copolímers de PEO- <i>b</i> -PCL: GPC, RMN i DSC	13
B.1 Multidetector gel permeation chromatography (GPC)	13
B.2 ¹ H-RMN	15
B.3 Differential scanning calorimetry (DSC)	18
C. Calibrat del sistema per l' <i>electrospinning</i>	25
D. Principis del mesurador de partícules	27
E. Microscopia òptica dels agregats de PEO- <i>b</i> -PCL obtinguts per diàlisi	35
F. Imatges de SEM dels copolímers fosos de PEO- <i>b</i> -PCL per <i>electrospinning</i>	37
F.1 Estudi del paràmetre temperatura	37
F.2 Estudi del paràmetre cabal	43
F.3 Estudi del paràmetre voltatge	46





Índex de figures

Figura B.1 Comparació de les corbes de GPC per a cada sèrie de monòmer i copolímers de PEO- <i>b</i> -PCL.....	14
Figura B.2 Espectre RMN del copolímer PEO ₂₀₀₀ - <i>b</i> -PCL ₄₀₀₀	15
Figura B.3 Espectre RMN del copolímer PEO ₂₀₀₀ - <i>b</i> -PCL ₆₀₀₀	16
Figura B.4 Espectre RMN del copolímer PEO ₂₀₀₀ - <i>b</i> -PCL ₈₀₀₀	16
Figura B.5 Espectre RMN del copolímer PEO ₅₀₀₀ - <i>b</i> -PCL ₁₀₀₀₀	17
Figura B.6 Espectre RMN del copolímer PEO ₅₀₀₀ - <i>b</i> -PCL ₁₅₀₀₀	17
Figura B.7 Espectre RMN del copolímer PEO ₅₀₀₀ - <i>b</i> -PCL ₂₀₀₀₀	18
Figura B.8 Termograma de DSC del copolímer PEO ₂₀₀₀ - <i>b</i> -PCL ₄₀₀₀	19
Figura B.9 Termograma de DSC del copolímer PEO ₂₀₀₀ - <i>b</i> -PCL ₆₀₀₀	19
Figura B.10 Termograma de DSC del copolímer PEO ₂₀₀₀ - <i>b</i> -PCL ₈₀₀₀	20
Figura B.11 Termograma de DSC del copolímer PEO ₅₀₀₀ - <i>b</i> -PCL ₁₀₀₀₀	20
Figura B.12 Termograma de DSC del copolímer PEO ₅₀₀₀ - <i>b</i> -PCL ₁₅₀₀₀	21
Figura B.13 Termograma de DSC del copolímer PEO ₅₀₀₀ - <i>b</i> -PCL ₂₀₀₀₀	21
Figura B.14 Termograma de DSC comparatiu entre els copolímers PEO ₂₀₀₀ - <i>b</i> -PCL ₄₀₀₀ , PEO ₂₀₀₀ - <i>b</i> -PCL ₆₀₀₀ i PEO ₂₀₀₀ - <i>b</i> -PCL ₈₀₀₀	22
Figura B.15 Termograma de DSC comparatiu entre els copolímers PEO ₅₀₀₀ - <i>b</i> -PCL ₁₀₀₀₀ , PEO ₅₀₀₀ - <i>b</i> -PCL ₁₅₀₀₀ i PEO ₅₀₀₀ - <i>b</i> -PCL ₂₀₀₀₀	22
Figura B.16 Termograma de DSC comparatiu de tots els copolímers de PEO- <i>b</i> -PCL.....	23
Figura C.1 Calibració per a la xeringa d'1 ml.....	25
Figura C.2 Calibració per a la xeringa de 2 ml.....	26
Figura C.3 Calibració del bany tèrmic.....	26
Figura E.1 Imatges de microscopia òptica de les dissolucions d'agregats. (a) PEO ₂₀₀₀ - <i>b</i> -PCL ₄₀₀₀ (IC); (b) PEO ₂₀₀₀ - <i>b</i> -PCL ₄₀₀₀ (IC); (c) PEO ₂₀₀₀ - <i>b</i> -PCL ₆₀₀₀ (IC); (d) PEO ₂₀₀₀ - <i>b</i> -PCL ₈₀₀₀ (IC); (e) PEO ₅₀₀₀ - <i>b</i> -PCL ₁₀₀₀₀ (IC); (f) PEO ₅₀₀₀ - <i>b</i> -PCL ₁₅₀₀₀ (DF) ; (g) PEO ₅₀₀₀ - <i>b</i> -PCL ₁₅₀₀₀ (DF) ; (h) PEO ₅₀₀₀ - <i>b</i> -PCL ₂₀₀₀₀ (IC).....	36
Figura F.1 Imatge de SEM d'una mostra del copolímer PEO ₂₀₀₀ - <i>b</i> -PCL ₈₀₀₀ representativa per a les mostres en el rang de temperatura 70-85 °C.....	39





Índex de taules

Taula A.1 Electrospinning de polímers fosos.....	7
Taula A.2 <i>Electrospinning</i> de dissolucions	8
Taula F.1 Imatges de SEM per a l'estudi de l'efecte de la temperatura en el copolímer PEO ₂₀₀₀ - <i>b</i> -PCL ₄₀₀₀	37
Taula F.2 Imatges de SEM per a l'estudi de l'efecte de la temperatura en el copolímer PEO ₂₀₀₀ - <i>b</i> -PCL ₆₀₀₀	38
Taula F.3 Imatges de SEM per a l'estudi de l'efecte de la temperatura en el copolímer PEO ₂₀₀₀ - <i>b</i> -PCL ₈₀₀₀	39
Taula F.4 Imatges de SEM per a l'estudi de l'efecte de la temperatura en el copolímer PEO ₅₀₀₀ - <i>b</i> -PCL ₁₀₀₀₀	40
Taula F.5 Imatges de SEM per a l'estudi de l'efecte de la temperatura en el copolímer PEO ₅₀₀₀ - <i>b</i> -PCL ₁₅₀₀₀	41
Taula F.6 Imatges de SEM per a l'estudi de l'efecte de la temperatura en el copolímer PEO ₅₀₀₀ - <i>b</i> -PCL ₂₀₀₀₀	42
Taula F.7 Imatges de SEM per a l'estudi de l'efecte del cabal en el copolímer PEO ₅₀₀₀ - <i>b</i> - PCL ₁₅₀₀₀	43
Taula F.8 Imatges de SEM per a l'estudi de l'efecte del cabal en el copolímer PEO ₅₀₀₀ - <i>b</i> - PCL ₂₀₀₀₀	44
Taula F.9 Imatges de SEM per a l'estudi de l'efecte del voltatge en el copolímer PEO ₅₀₀₀ - <i>b</i> - PCL ₁₅₀₀₀ a 0.05 ml/h	46
Taula F.10 Imatges de SEM per a l'estudi de l'efecte del voltatge en el copolímer PEO ₅₀₀₀ - <i>b</i> - PCL ₁₅₀₀₀ a 0.1 ml/h	47
Taula F.11 Imatges de SEM per a l'estudi de l'efecte del voltatge en el copolímer PEO ₅₀₀₀ - <i>b</i> - PCL ₁₅₀₀₀ a 0.2 ml/h	47
Taula F.12 Imatges de SEM per a l'estudi de l'efecte del voltatge en el copolímer PEO ₅₀₀₀ - <i>b</i> - PCL ₂₀₀₀₀ a 0.05 ml/h	48
Taula F.13 Imatges de SEM per a l'estudi de l'efecte del voltatge en el copolímer PEO ₅₀₀₀ - <i>b</i> - PCL ₂₀₀₀₀ a 0.1 ml/h	49
Taula F.14 Imatges de SEM per a l'estudi de l'efecte del voltatge en el copolímer PEO ₅₀₀₀ - <i>b</i> - PCL ₂₀₀₀₀ a 0.2 ml/h	49





A. Electrospinning de dissolucions i polímers fosos

Taula A.1 Electrospinning de polímers fosos

No.	Polymer	Material details	Processing temp. (°C)
1	Polyethylene, PE	HDPE, $M_w = 1.35 \times 10^5$ [95,97] HDPE [120] [121]	200–220
2	Polypropylene, PP	Isotactic-PP, MI = 0.5 [95] [120] [121]	220–240
3	Nylon 12, PA-12	$M_w = 3.5 \times 10^4$ [96]	220
4	Polyethylene terephthalate, PET	[120] [121] $M_w = 4.6 \times 10^4$ [87]	270
5	Polyethylene naphthalate, PEN	[121] $M_w = 4.8 \times 10^4$ [87]	290
6	PET/PEN blends	75/25, 25/75 (wt.%), [87]	290



Taula A.2 *Electrospinning* de dissolucions

Polymers to have been electrospun in solution form					
No.	Polymer	Details ^a	Solvent	Concentration	Perspective applications
1	Nylon6,6, PA-6,6	[127]	Formic acid	10 wt. %	Protective clothing
2	Polyurethanes, PU	[127] [152]	Dimethyl formamide Dimethylformamide	10 wt. % 10 wt. %	Protective clothing Electret filter
3	Polybenzimidazole, PBI	[127] [88]	Dimethyl acetamide	10 wt. %	Protective clothing nanofiber reinforced composites
4	Polycarbonate, PC	[127] [9] $M_w = 60,000$ [114] $MI = 8-10$ g/10 min [92] [152]	Dimethyl formamide:tetrahydrofuran (1:1) Dichloromethane Chloroform, tetrahydrofuran Dimethylformamide:tetrahydrofuran (1:1) Dimethylformamide:tetrahydrofuran (1:1)	10 wt. % 15 wt. % 14–15 wt. % 20 wt. %	Protective clothing Sensor, filter Electret filter
6	Polyacrylonitrile, PAN	[156] [157] [40] [125] [158]	Dimethyl formamide Dimethyl formamide Dimethyl formamide Dimethyl formamide	600 mgPAN/10-5m3 dimethylformamide 15 wt. %	Carbon nanofiber
7	Polyvinil alcohol, PVA	$M_n = 65,000$ [34] [129] $M_n = 150,000$ [115]	Distilled water Distilled water	8–16 wt. % 4–10 wt. % 1–10 wt. %	
8	Polylactic acid, PLA	poly(D, L-lactic acid) $M_w = 109,000$ [168] poly(L-lactic acid) $M_w = 100,000$ [168] poly(L-lactic acid) $M_n = 150,000$ g/mol [10] $M_w = 205$ kDa [84] [21]	Dimethyl formamide Methylene chloride and dimethyl formamide Dichloromethane Dichloromethane	5 wt. % 14 wt. % 14 wt. % 14 wt. %	Membrane for prevention of surry induced-adhesion Sama as above Sensor, Filter Drug delivery system Drug delivery system Drug delivery system
9	Polyethylene-co-vinyl acetate, PEVA	$M_w = 60.4$ kDa [84]			
10	PEVA/PLA	PEVA/PLA = 50/50 [84]			



Table 1 (continued)

No.	Polymer	Details ^a	Solvent	Concentration	Perspective applications
18	Polyethylene Terephthalate, PET	$M_w = 10,000\text{--}20,000$ g/mol [122] [158]	Dichloromethane and trifluoroacetic	4 wt. %	
19	polyacrylic acid-polypyrrole methanol, PAA-PM	$M_w = 50,000$ g/mol [158]	Dichloromethane:trifluoroacetic acid (1:1) Dimethyl formamide	12–18 wt. %	Optical sensor
20	Polystyrene, PS	$M_w = 190,000$ [114] $M = 200$ kDa [81] [129] [141] $M = 280,000$ [90] $M_w = 280,000$ [151] $M_w = 280,000/M_w = 28,000$: 90/1 [151] $M_w = 280,000/M_w = 28,000$: 50/50 [151] $M_w = 280,000/M_w = 2,430$:90/10 [151]	Tetrahydrofuran, dimethylformamide, CS ₂ (carbon disulfide), toluene, Methyl ethyl ketone Chloroform, dimethylformamide Tetrahydrofuran Dimethylformamide Tetrahydrofuran Tetrahydrofuran Tetrahydrofuran Tetrahydrofuran	18–35 wt. % 8% 2.5–10.7% 25 wt. % 30 wt. % 15 wt. % 15 wt. % 15 wt. % 15 wt. %	Enzymatic biotransformation (Flat ribbons) Catalyst, filter Catalyst, filter Catalyst, filter Catalyst, filter
21	Polymethacrylate, PMMA	$M_w = 540,000$ [114]	Tetrahydrofuran, acetone, chloroform		
22	Polyamide, PA	[66]	Dimethylacetamide		Glass fiber filter media
23	Silk/PEO blend	$M_w(\text{PEO}) = 900,000$ g/mol [82]	Silk aqueous solutions	4.8–8.8 wt. %	Biomaterial scaffolds
24	poly vinyl phenol, PVP	$M_w = 20,000$, 100,000 [83]	Tetrahydrofuran	20, 60% (wt./vol.)	Antimicrobial agent
25	Polyvinylchloride, PVC	[100,101]	Tetrahydrofuran/dimethylformamide = 100/0, 80/20, 60/40, 50/50, 40/60, 20/80, 0/100 (vol.%)	10–15 wt. %	
26	Cellulose acetate, CA	[105]	Acetone, acetic acid, dimethylacetamide	12.5–20%	Membrane
27	Mixture of PAA-PM (polyacrylic acid – poly (pyrene methanol)) and polyurethane	[154]	Dimethylformamide	26 wt. %	Optical sensor
28	Polyvinil alcohol (PVA)/Silica,	PVA: $M_n = 86,000$, silica content (wt. %): 0, 22, 34, 40, 49, 59 [132]	Distilled water		



Table 1 (continued)

No.	Polymer	Details ^a	Solvent	Concentration	Perspective applications
29	Polyacrylamide, PAAm	$M_n = 5,000,000$ [115]		1–10 wt. %	
30	PLGA	PLGA(PLA/PGA) = (85/15) [102]	Tetrahydrofuran:dimethylformamide (1:1)	1 g/20 ml	Scaffold for tissue engineering
31	Collagen	[113]	Hexafluoro-2-propanol		Scaffold for tissue engineering
32	Polycaprolactone, PCL	[125]	Chloroform:methanol (3:1) toluene:methanol (1:1), and dichloromethane:methanol (3:1)		
33	Poly(2-hydroxyethyl methacrylate), HEMA	$M = 200,000$ [90]	Ethanol:formic acid (1:1), ethanol	12, 20 wt. % / 8, 16, 20 wt. %	(Flat ribbons)
34	Poly(vinylidene fluoride), PVDF	$M = 107,000$ [90]	Dimethylformamide:dimethylacetamide (1/1)	20 wt. %	(Flat ribbons)
35	Polyether imide, PEI	[90]	Hexafluoro-2-propanol	10 wt. %	(Flat ribbons)
36	Polyethylene gricol, PEG	$M = 10\text{ K}$ [158]	Chloroform	0.5–30 wt. %	
37	nylon-4,6, PA-4,6	[7]	Formic acid	10 wt. %	Transparent composite
38	Poly(ferrocenyldimethylsilane), PFDMS	$M_w = 87,000$ g/mol [22]	Tetrahydrofuran:dimethylformamide (9:1)	30 wt. %	
39	Nylon6 (PA-6) /montmorillonite (Mt)	M_t content = 7.5 wt. % [50]	Hexa-fluoro-isopropanol (HFIP), HFIP/dimethylformamide: 95/5 (wt%)	10 wt. %	
40	poly(ethylene-co-vinyl alcohol)	Vinyl alcohol repeat unit: 56–71 mol% [85]	Isopropanol/water: 70/30 (%v/v)	2.5–20%w/v	Biomedical
41	Polyacrylonitrile (PAN) / TiO ₂	[168]			
42	Polycaprolactone (PCL) / metal	Metals: gold, ZnO, [124]			Photovoltaic and conductive polymers
43	Polyvinyl pyrrolidone, PVP	[26]			ZnO: cosmetic use
44	Polymetha-phenylene isophthalamide	[124]			

^a Details possibly include: (a) reference, (b) molecular weight, and (c) content of each polymer in co-polymer/blend/composite.



B. Caracterització dels copolímers de PEO-*b*-PCL: GPC, RMN i DSC

B.1 Multidetector gel permeation chromatography (GPC)

Les corbes de GPC s'han obtingut en un aparell de columna mixta tipus D (300 mm x 8 mm) de Polymer Laboratories Ltd. Per a realitzar una calibració universal de l'aparell s'han emprat estàndards de petits intervals de pes molecular de poliestirè (pesos moleculars entre 580 g/mol i 9995600 g/mol). Amb el programa Win-GPC 5.1.0 s'han extret de les corbes els pesos moleculars (M_n i M_w) i la polidispersitat. Els pesos moleculars no han estat corregits i són en g/mol. Els polímers s'han dissolt en THF a una concentració de 10mg/ml i s'han injectat 100 µl de cada dissolució per a obtenir la mesura.

Sistema THF:

Dissolvent:	THF destil·lat amb sodi i benzofenona amb una dissolució de 250 mg/l de 2,6-Di- <i>tert</i> -butil-4-metilfenol
Cabal de circulació:	1 ml/min
Autosampler:	Schambeck S5200
Bomba HPLC:	Waters Millipore 510
Detector UV:	ERC-7215
Detector IR:	ERC-7515A
Temperatura de la columna:	Temperatura ambient
Combinació de columnes:	MZ Gel (d = 8.0 mm, l = 50 mm) 10^3 Å (Precolumna) MZ Gel (d = 8.0 mm, l = 300 mm) 10^3 Å MZ Gel (d = 8.0 mm, l = 50 mm) 10^3 Å MZ Gel (d = 8.0 mm, l = 50 mm) 10^4 Å MZ Gel (d = 8.0 mm, l = 50 mm) 10^5 Å



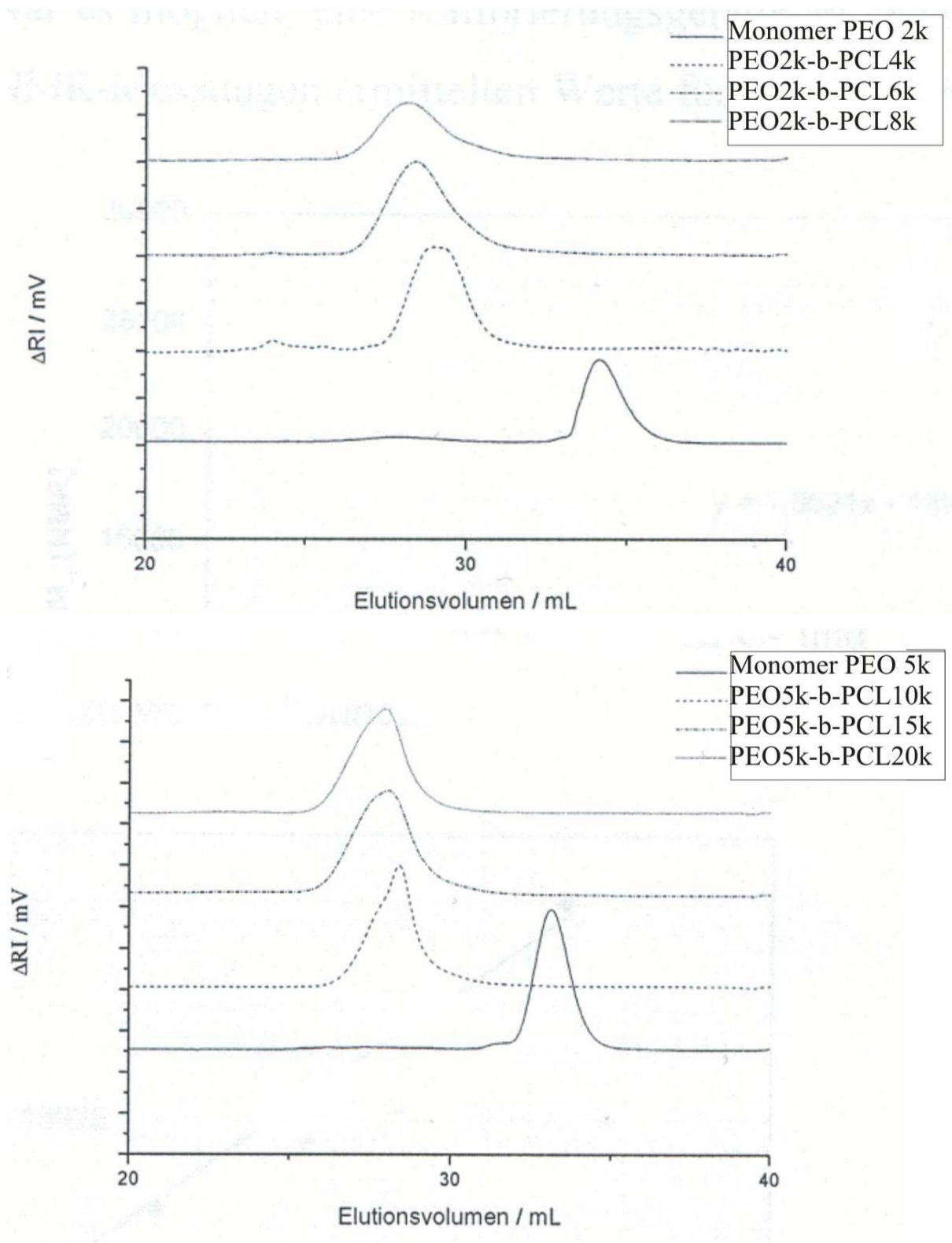


Figura B.1 Comparació de les corbes de GPC per a cada sèrie de monòmer i copolímers de PEO-*b*-PCL



B.2 ^1H -RMN

Els espectres RMN s'han obtingut amb un aparell Varian VXR 300:

Varian Gemini 300 (300 MHz)

Varian Inova 400 (400 MHz)

Varian Unity 500 (500 MHz)

Estàndard intern: Tetrametilsilà

L'acoplament químic δ està referenciat en ppm respecte el tetrametilsilà. Les unitats de les constants d'acoplaments són $[J] = \text{Hz}$.

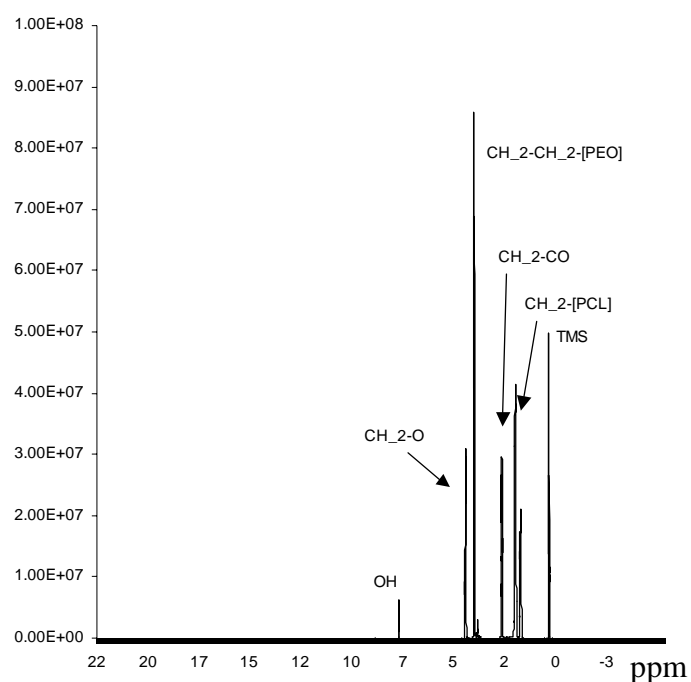


Figura B.2 Espectre RMN del copolímer $\text{PEO}_{2000}\text{-b-PCL}_{4000}$



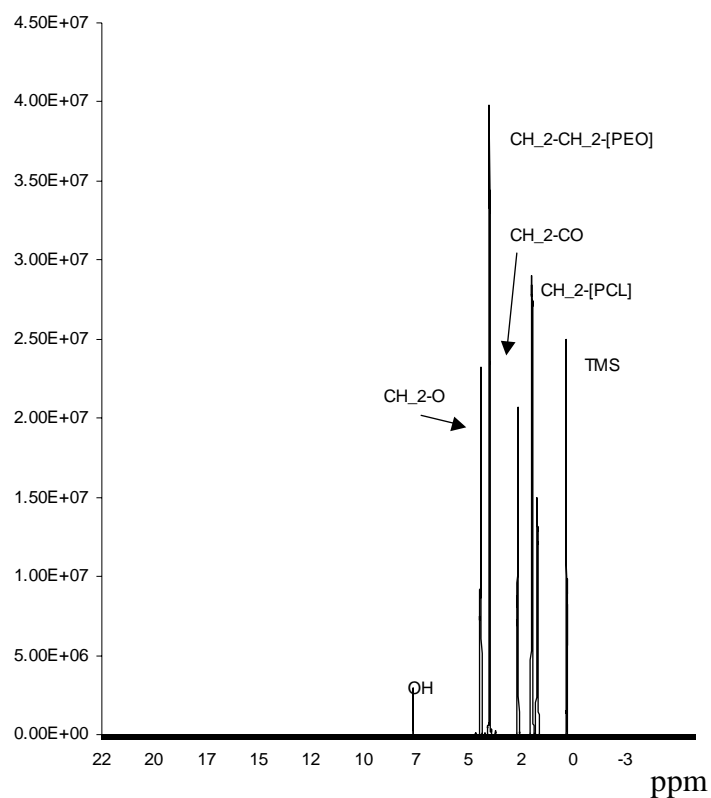


Figura B.3 Espectre RMN del copolímer $\text{PEO}_{2000}\text{-b-PCL}_{6000}$

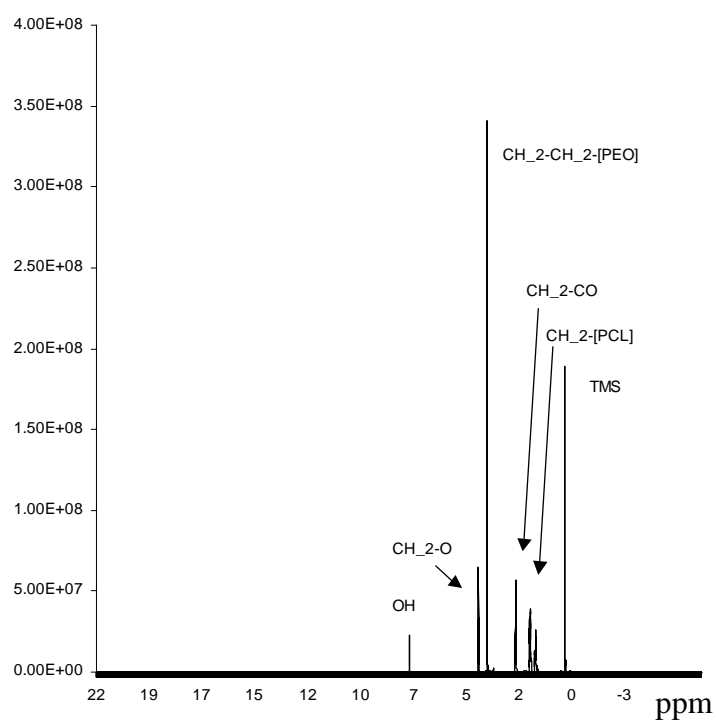
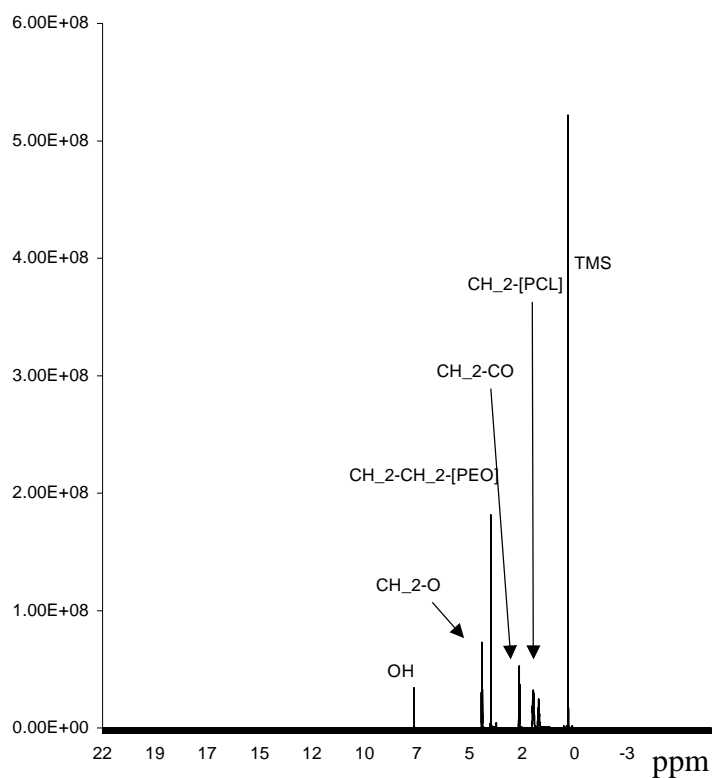
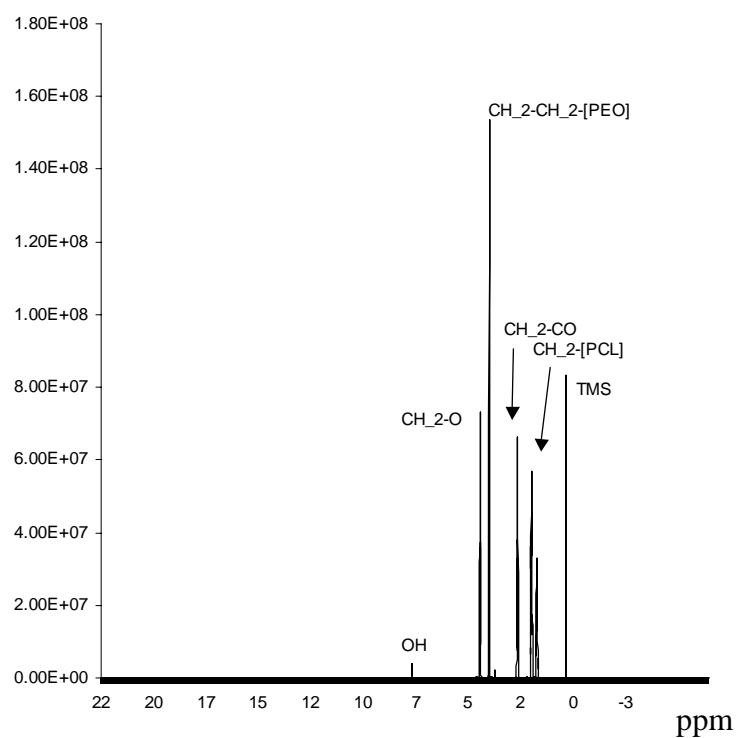


Figura B.4 Espectre RMN del copolímer $\text{PEO}_{2000}\text{-b-PCL}_{8000}$



Figura B.5 Espectre RMN del copolímer PEO₅₀₀₀-b-PCL₁₀₀₀₀Figura B.6 Espectre RMN del copolímer PEO₅₀₀₀-b-PCL₁₅₀₀₀

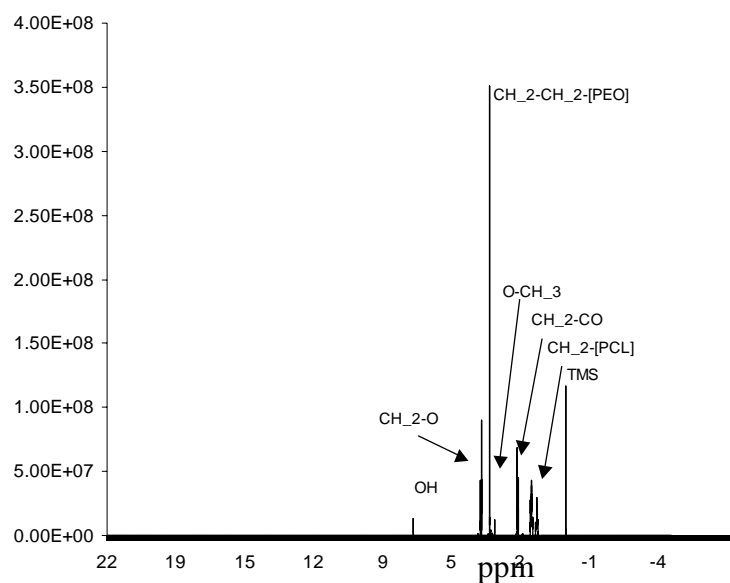


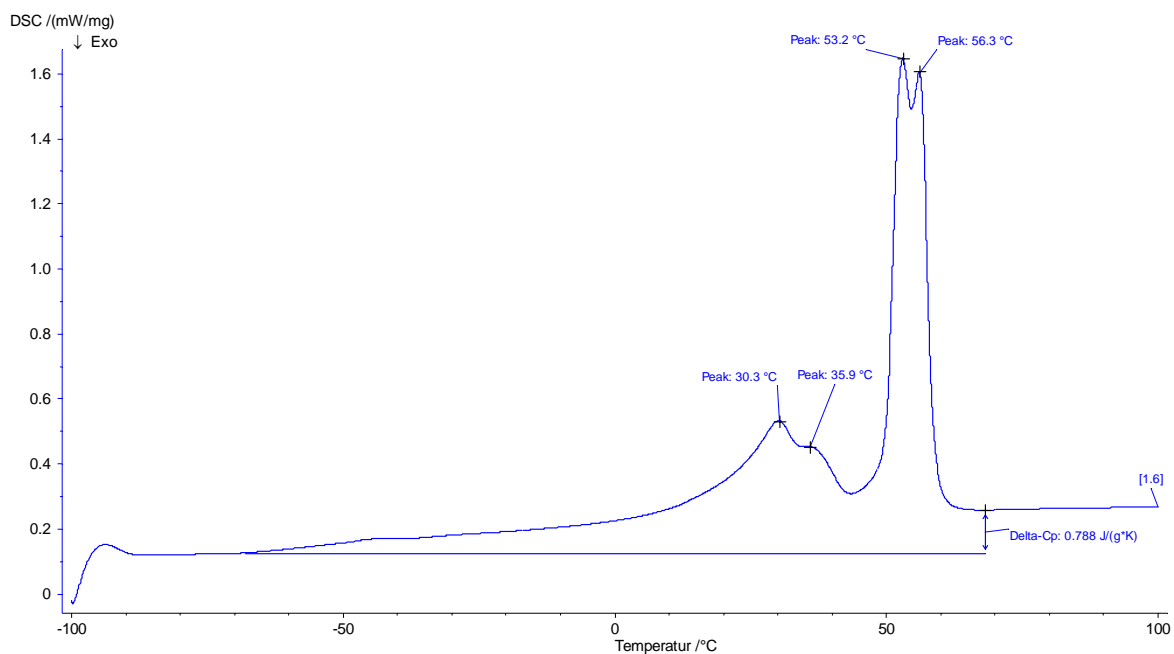
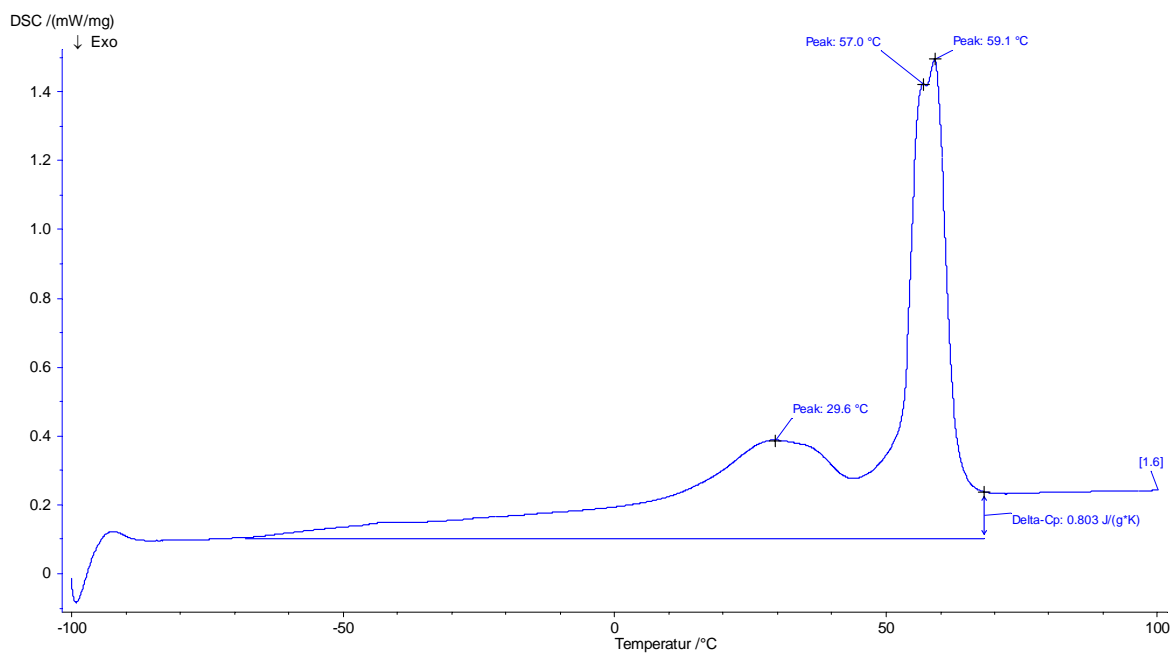
Figura B.7 Espectre RMN del copolímer PEO₅₀₀₀-b-PCL₂₀₀₀₀

B.3 Differential scanning calorimetry (DSC)

Els termogrames de DSC s'han realitzat en un aparell Perkin Elmer DSC7 equipat amb una cel·la de baixa temperatura i un processador TA7. Els intervals de temperatura s'han ajustat al rang d'interès. S'han introduït uns 5 mg de mostra per a cada mesura. Les transicions han estat avaluades utilitzant el software Pyris Software (Perkin Elmer). Les mostres han estat a dins de l'aparell permanentment sota atmosfera de N₂.

Els termogrames de DSC mostrats són els corresponents a la sisena etapa. En aquesta etapa s'ha programat escalfar entre -100°C i 100°C a 10°C / min.



Figura B.8 Termograma de DSC del copolímer PEO₂₀₀₀-*b*-PCL₄₀₀₀Figura B.9 Termograma de DSC del copolímer PEO₂₀₀₀-*b*-PCL₆₀₀₀

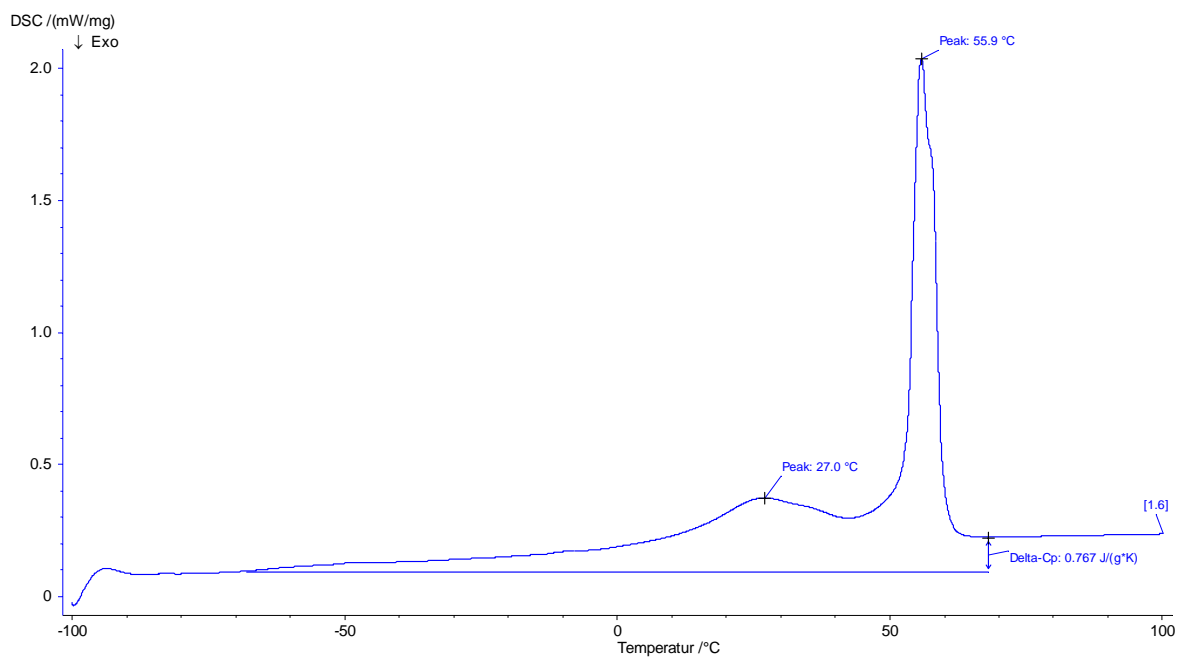


Figura B.10 Termograma de DSC del copolímer PEO₂₀₀₀-*b*-PCL₈₀₀₀

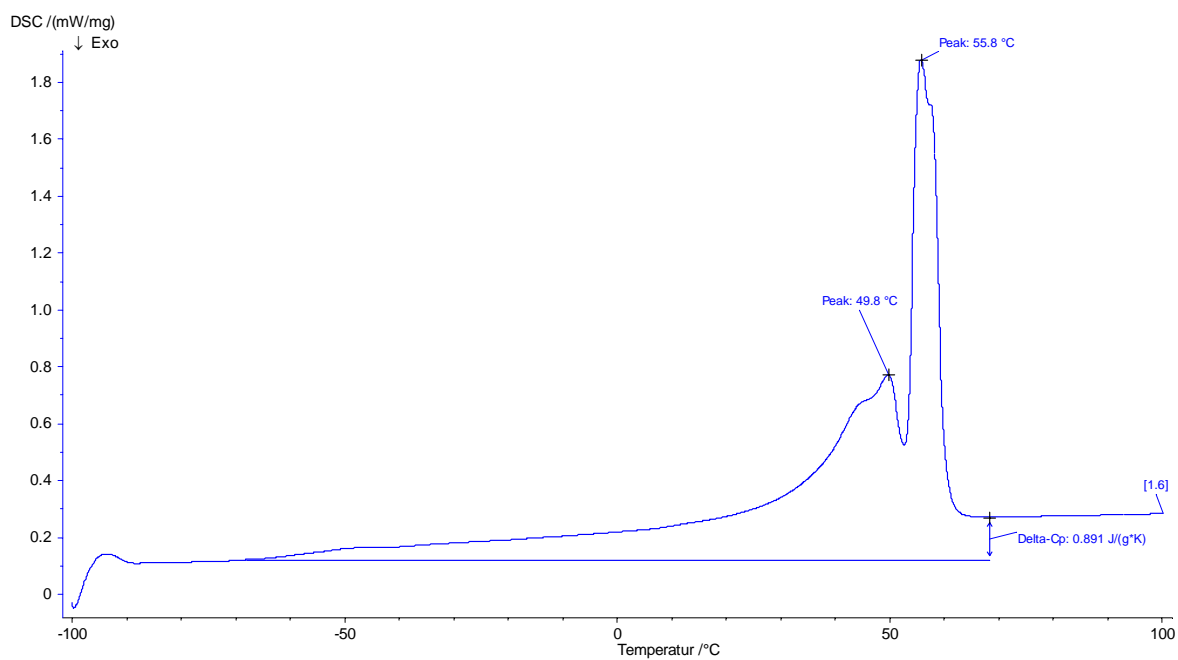
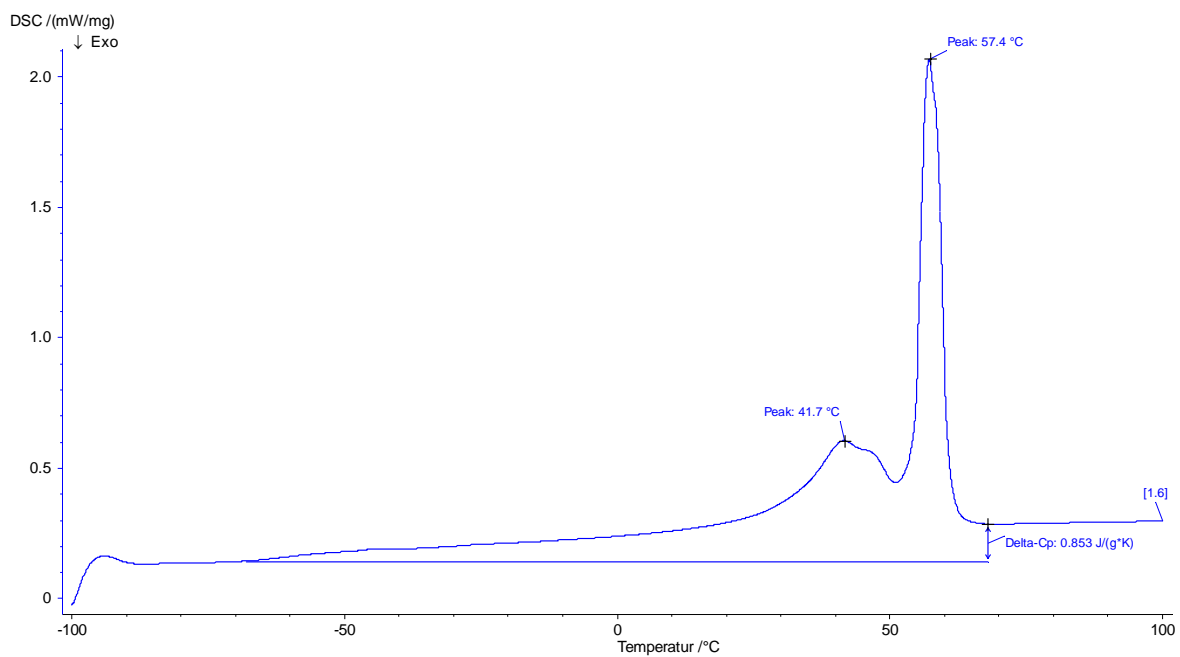
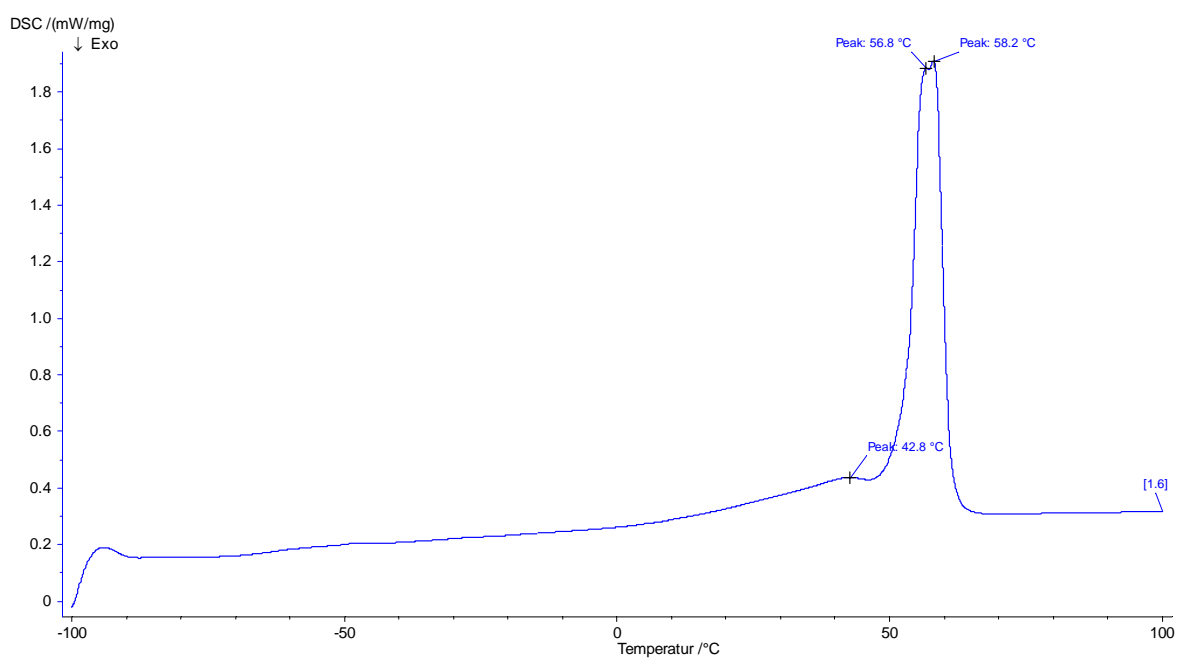


Figura B.11 Termograma de DSC del copolímer PEO₅₀₀₀-*b*-PCL₁₀₀₀₀



Figura B.12 Termograma de DSC del copolímer PEO₅₀₀₀-*b*-PCL₁₅₀₀₀Figura B.13 Termograma de DSC del copolímer PEO₅₀₀₀-*b*-PCL₂₀₀₀₀

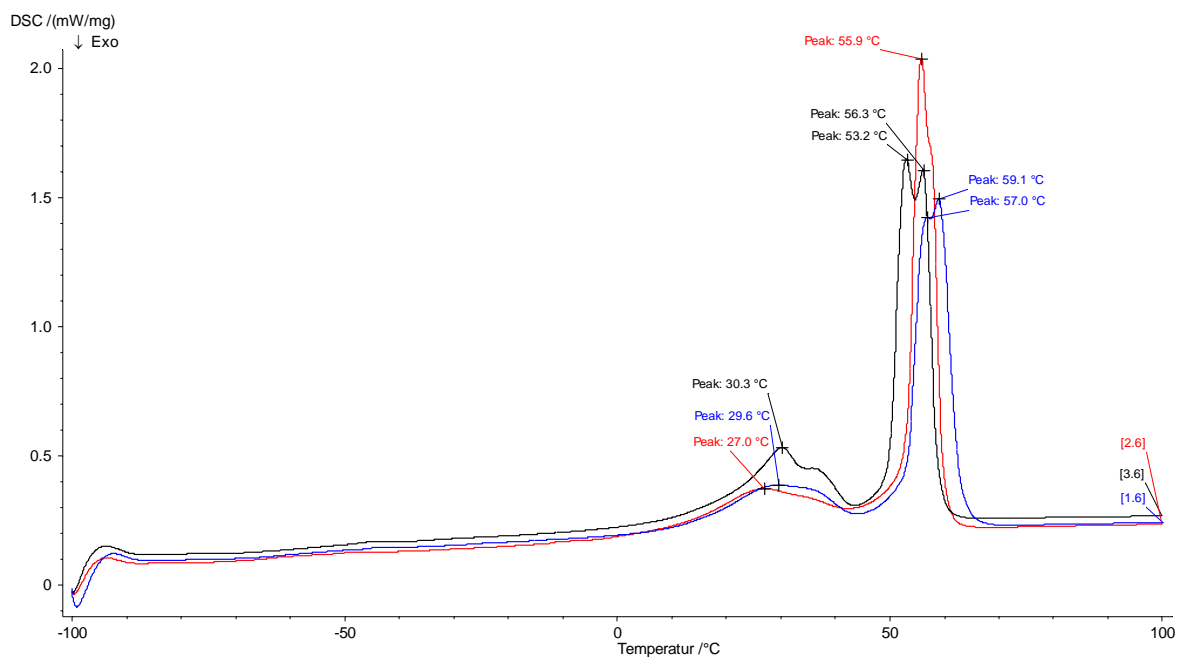


Figura B.14 Termograma de DSC comparatiu entre els copolímers $\text{PEO}_{2000}\text{-}b\text{-PCL}_{4000}$, $\text{PEO}_{2000}\text{-}b\text{-PCL}_{6000}$ i $\text{PEO}_{2000}\text{-}b\text{-PCL}_{8000}$

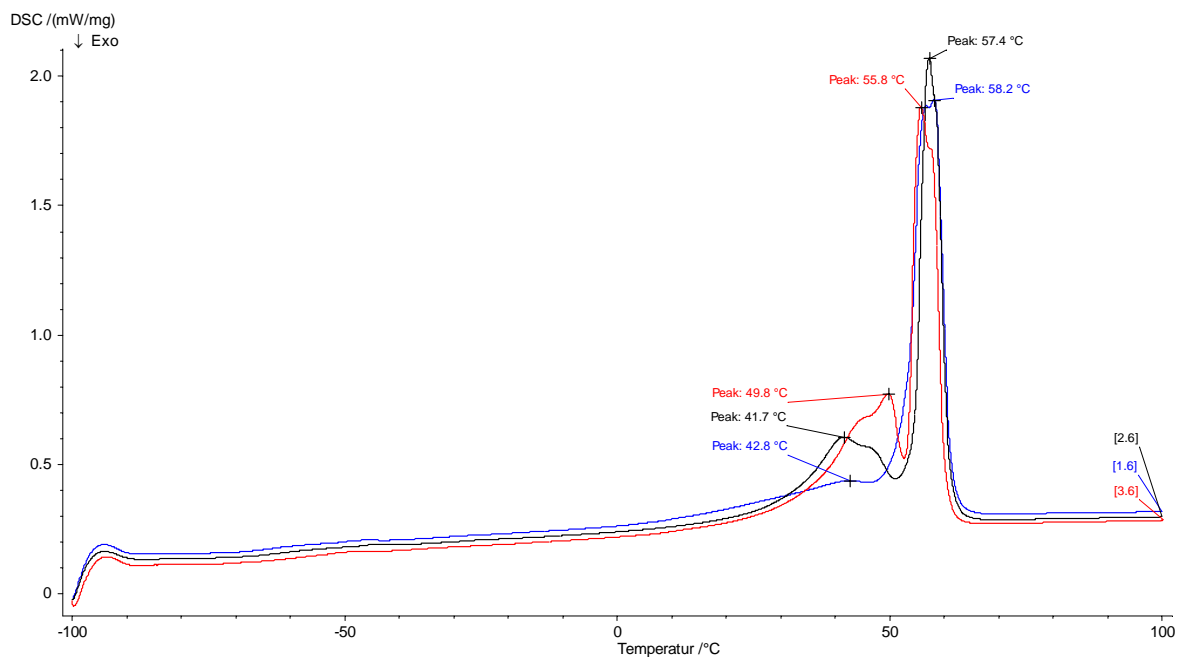
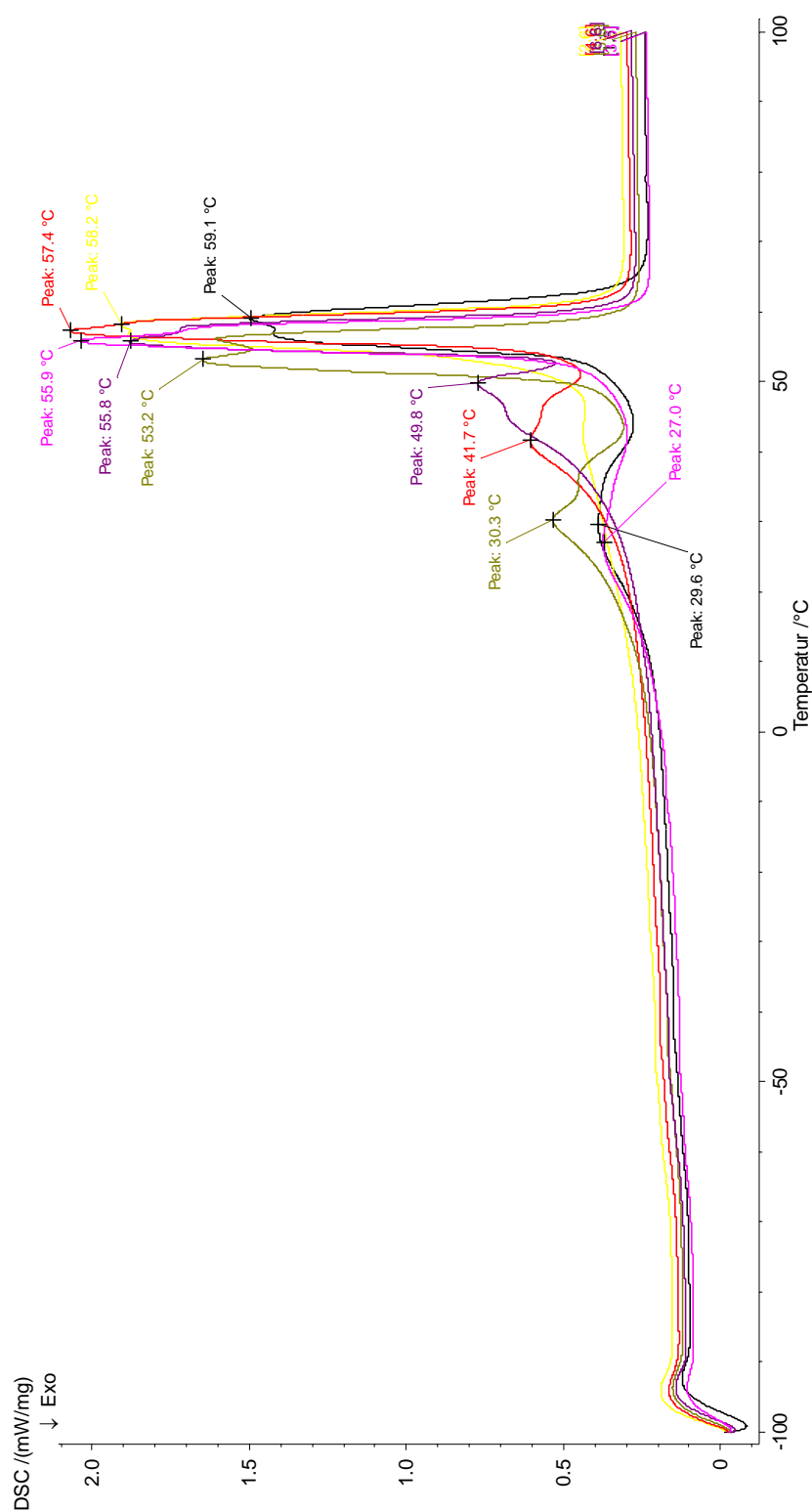


Figura B.15 Termograma de DSC comparatiu entre els copolímers $\text{PEO}_{5000}\text{-}b\text{-PCL}_{10000}$, $\text{PEO}_{5000}\text{-}b\text{-PCL}_{15000}$ i $\text{PEO}_{5000}\text{-}b\text{-PCL}_{20000}$



Figura B.16 Termograma de DSC comparatiu de tots els copolímers de PEO-*b*-PCL



C. Calibrat dels sistema per l'*electrospinning*

La calibració de la bomba d'injecció i del bany tèrmic han sigut necessàries a fi de poder fixar correctament les variables corresponents quan s'ha procedit a l'*electrospinning*. Les corbes de calibrat s'han obtingut per a les xeringues d'1 i 2 ml perquè són les que s'han emprat amb més freqüència (respectivament Figura C.1 i C.2). Per a la calibració, s'ha deixat funcionar la bomba d'injecció durant 1 h per a cada mesura, omplint una segona xeringa (unida per un connector).

També s'ha fet la calibració del bany tèrmic col·locant un termòmetre a una altura mitja a dins del condensador (Figura C.3). S'han pres mesures cada 5 °C entre 60 i 90°C deixant que la temperatura s'estabilitzés durant 15 minuts.

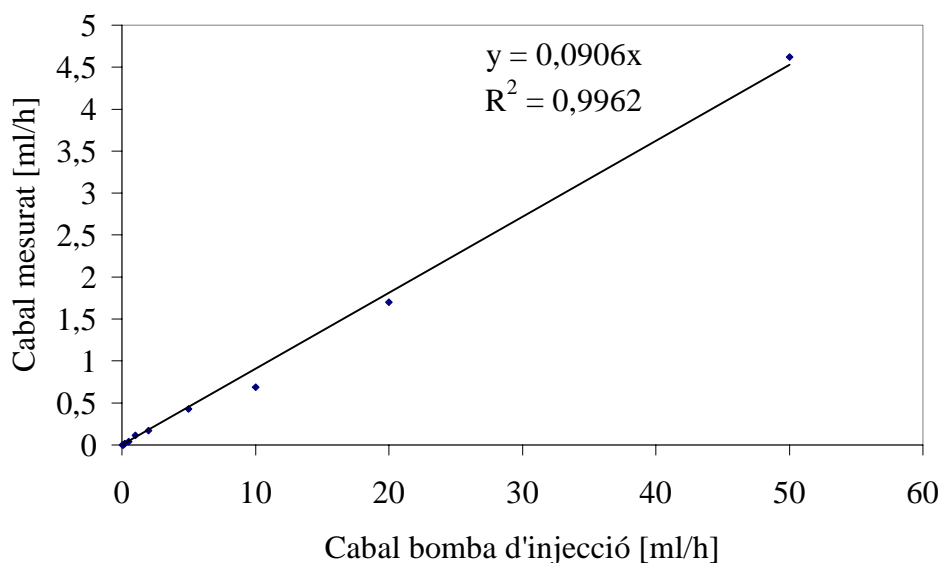


Figura C.1 Calibració per a la xeringa d'1 ml



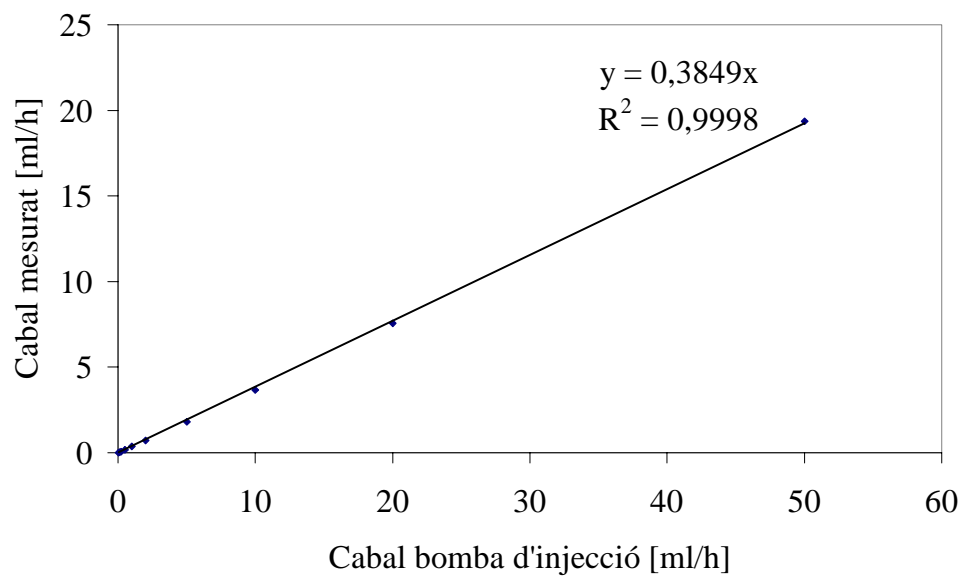


Figura C.2 Calibració per a la xeringa de 2 ml

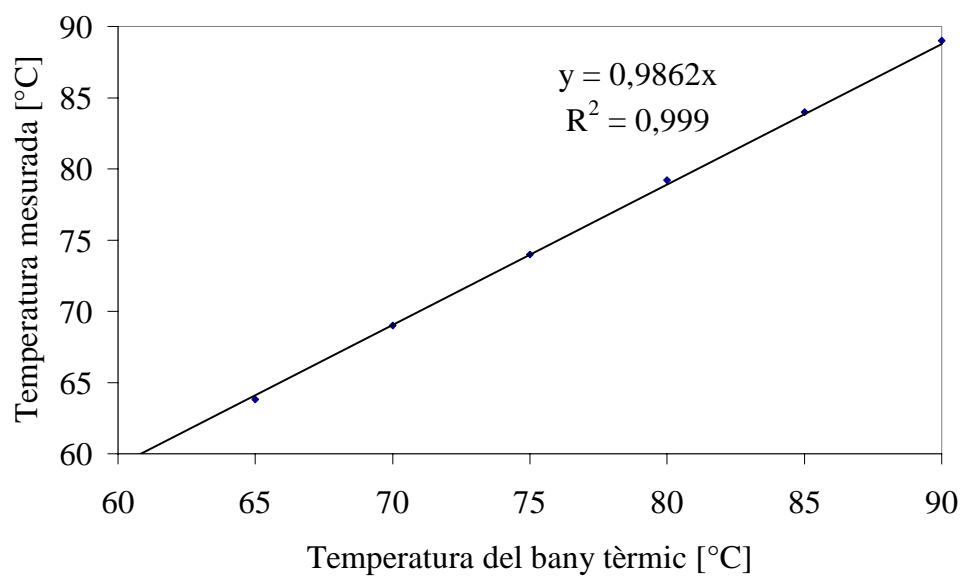


Figura C.3 Calibració del bany tèrmic



D. Principis del mesurador de partícules

BASIC PRINCIPLES OF PARTICLE SIZE ANALYSIS

Written by Dr. Alan Rawle, Malvern Instruments Limited, Enigma Business Park, Grovewood Road, Malvern, Worcestershire, WR14 1XZ, UK. Tel: +44 (0)1684 892456 Fax: +44 (0)1684 892789

What is a Particle?

This may seem a fairly stupid question to ask! However, it is fundamental in order to understand the results which come from various particle size analysis techniques. Dispersion processes and the shape of materials makes particle size analysis a more complex matter than it first appears.

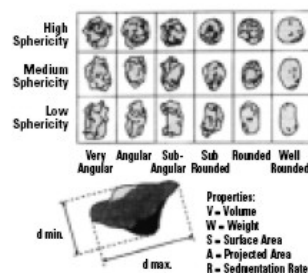


Figure 1

The Particle size conundrum

Imagine that I give you a matchbox and a ruler and ask you to tell me the size of it. You may reply saying that the matchbox is 20 x 10 x 5mm. You cannot correctly say "the matchbox is 20mm" as this is only one aspect of its size. So it is not possible for you to describe the 3-dimensional matchbox with one unique number. Obviously the situation is more difficult for a complex shape like a grain of sand or a pigment particle in a can of paint. If I am a Q.A. Manager, I will want one number only to describe my particles – I will need to know if the average size has increased or decreased since the last production run, for example. This is the basic problem of particle size analysis – how do we describe a 3-dimensional object with one number only?

Figure 1 shows some grains of sand. What size are they?

The equivalent sphere

There is only one shape that can be described by one unique number and that is the sphere. If we say that we have a 50µ sphere, this describes it exactly. We cannot do the same even for a cube where 50µ may refer to an

edge or to a diagonal. With our matchbox there are a number of properties of it that can be described by one number. For example the weight is a single unique number as is the volume and surface area. So if we have a technique that measures the weight of the matchbox, we can then convert this weight into the weight of a sphere, remembering that...

$$\text{weight} = \frac{4}{3} \pi r^3 \cdot \rho$$

and calculate one unique number (2r) for the diameter of the sphere of the same weight as our matchbox. This is the equivalent sphere theory. We measure some property of our particle and assume that this refers to a sphere, hence deriving our one unique number (the diameter of this sphere) to describe our particle. This ensures that we do not have to describe our 3-D particles with three or more numbers which although more accurate is inconvenient for management purposes.

We can see that this can produce some interesting effects depending on the shape of the object and this is illustrated by the example of equivalent spheres of cylinders (Fig. 2). However, if our cylinder changes shape or size then the volume/weight will change and we will at least be able to say that it has got larger/smaller etc. with our equivalent sphere model.

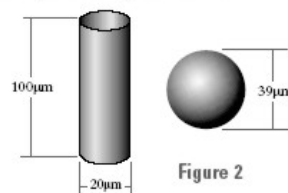


Figure 2

Equivalent spherical diameter of cylinder 100 x 20µm

Imagine a cylinder of diameter $D_1 = 20\mu\text{m}$ (i.e. $r=10\mu\text{m}$) and height $100\mu\text{m}$.

There is a sphere of diameter, D_2 which has an equivalent volume to the cylinder. We can calculate this diameter as follows:

Volume of cylinder =

$$\pi r^2 h = 10000 \pi (\mu\text{m}^3)$$

Volume of sphere =

$$\frac{4}{3} \pi X^3$$

Where X is equivalent volume radius.

$$\therefore X = \sqrt[3]{\frac{3V}{4\pi}} = 0.62 \sqrt[3]{V}$$

$$X = \sqrt[3]{\frac{30000\pi}{4\pi}} = \sqrt[3]{7500} = 19.5\mu\text{m}$$

$$\therefore D_2 = 39.1\mu\text{m}$$

The volume equivalent spherical diameter for a cylinder of 100µm height and 20µm in diameter is around 40µm. The table below indicates equivalent spherical diameters of cylinders of various ratios. The last line may be typical of a large clay particle which is discshaped. It would appear to be 20µm in diameter, but as it is only 0.2µm in thickness, normally we would not consider this dimension. On an instrument which measures the volume of the particle we would get an answer around 5µm. Hence the possibility for disputing answers that different techniques give!

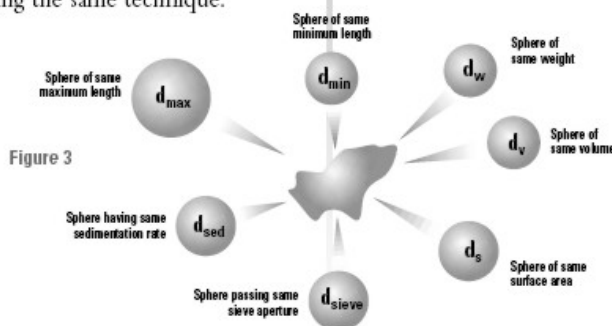
Note also that all these cylinders will appear the same size to a sieve, of say 25µm where it will be stated that "all material is smaller than 25µm". With laser diffraction these 'cylinders' will be seen to be different because they possess different values.



Size of cylinder		Aspect Ratio	Equivalent Spherical Diameter
Height	Diam.		
20	20	1:1	22.9
40	20	2:1	28.8
100	20	5:1	39.1
200	20	10:1	49.3
400	20	20:1	62.1
10	20	0.5:1	18.2
4	20	0.2:1	13.4
2	20	0.1:1	10.6

Different techniques

Clearly if we look at our particle under the microscope we are looking at some 2-D projection of it and there are a number of diameters that we can measure to characterise our particle. If we take the maximum length of the particle and use this as our size, then we are really saying that our particle is a sphere of this maximum dimension. Likewise, if we use the minimum diameter or some other quantity like Feret's diameter, this will give us another answer as to the size of our particle. Hence we must be aware that each characterisation technique will measure a different property of a particle (max. length, min. length, volume, surface area etc.) and therefore will give a different answer from another technique which measures an alternative dimension. Figure 3 shows some of the different answers possible for a single grain of sand. Each technique is not wrong – they are all right – it is just that a different property of the particle is being measured. It is like you measure your matchbox with a cm ruler and I measure with an inch ruler (and you measure the length and I measure the width!). Thus we can only seriously compare measurements on a powder by using the same technique.



This also means that there cannot be anything like particle size standard for particles like grains of sand. Standards must be spherical for comparison between techniques. However we can have a particle size standard for a particular technique and this should allow comparison between instruments which use that technique.

D[4,3]etc

Imagine three spheres of diameters 1,2,3 units. What is the average size of these three spheres? On first reflection we may say 2.00. How have we got this answer? We have summed all the diameters

$$(\sum d = 1 + 2 + 3)$$

and divided by the number of particles ($n=3$). This is a number mean, (more accurately a number length mean), because the number of the particles appears in the equation: Mean diameter =

$$\frac{1 + 2 + 3}{3} = 2.00 = \frac{\sum d}{n}$$

In mathematical terms this is called the $D[1,0]$ because the diameter terms on the top of the equation are to the power of (d^1) and there are no diameter terms (d^0) on the bottom equation.

However imagine that I am a catalyst engineer. I will want to compare these spheres on the basis of surface area because the higher the surface area, the higher the activity of the catalyst. The surface area of a sphere is $4\pi r^2$. Therefore to compare on basis of surface area we must square the diameters, divide by the number of particles, and take the square root

to get back to a mean diameter:

$$\sqrt{\frac{(1^2 + 2^2 + 3^2)}{3}} = 2.16 = \frac{\sum d^2}{n}$$

This is again a number mean (number-surface mean) because the number of particles appear on the bottom of the equation. We have summed the squares of the diameter so in mathematical terms this is called the $D[2,0]$ – diameter terms squared on the top, no diameter terms on the bottom.

If I am a chemical engineer I will want to compare the spheres on the basis of weight. Remembering that the weight of a sphere is:

$$\frac{4}{3} \pi r^3 \cdot \rho$$

then we must cube the diameters, divide by the number of particles and take a cube root to get back to a mean diameter:

$$\sqrt[3]{\frac{(1^3 + 2^3 + 3^3)}{3}} = 2.29 = \frac{\sum d^3}{n}$$

Again this is a number mean (number-volume or number-weight mean) because the number of particles appears in the equation. In mathematical terms this can be seen to be $D[3,0]$.

The main problem with the simple means, $D[1,0]$, $D[2,0]$, $D[3,0]$, is that the number of particles is inherent in the formulae. This gives rise to the need to count large numbers of particles. Particle counting is normally only carried out when the numbers are very low (in the ppm or ppb regions) in applications such as contamination, control and cleanliness. A simple calculation shows that in 1g of silica (density 2.5) then there would be around 760×10^9 particles if they were all $1\mu\text{m}$ in size.

Hence the concept of Moment Means needs to be introduced and this is usually where some confusion can arise. The two most important moment means are the following:

- $D[3,2]$ – Surface Area Moment Mean – Sauter Mean Diameter
- $D[4,3]$ – Volume or Mass Moment Mean – De Brouckere Mean Diameter



These means are analogous to moments of inertia and introduce another linear term in diameter (i.e. surface area has a d^3 dependence and volume or mass a d^4 dependence as below):

$$D[4,3] = \frac{1^4 + 2^4 + 3^4}{1^3 + 2^3 + 3^3} = 2.72 = \frac{\sum d^4}{\sum d^3}$$

$$D[3,2] = \frac{1^3 + 2^3 + 3^3}{1^2 + 2^2 + 3^2} = 2.57 = \frac{\sum d^3}{\sum d^2}$$

These formulae indicate around which central point of the frequency the (surface area or volume/mass) distribution would rotate. They are, in effect, centres of gravity of the respective distributions. The advantage of this method of calculation is obvious – the formulae do not contain the numbers of particles and therefore calculations of the means and distributions do not require knowledge of the number of particles involved. Laser diffraction initially calculates a distribution based around volume terms and this is why the $D[4,3]$ is reported in a prominent manner.

Different techniques give different means.

If we use an electron microscope to measure our particles it is likely that we will measure the diameters with a graticule, add them up and divide by the number of particles to get a mean result. We can see that we are generating the $D[1,0]$ number-length mean by this technique. If we have access to some form of image analysis then the area of each particle is measured and divided by the number of particles – the $D[2,0]$ is generated. If we have a technique like electrozone sensing, we will measure the volume of each particle and divide by the number of particles – a $D[3,0]$ is generated.

Laser diffraction can generate the $D[4,3]$ or equivalent volume mean. This is identical to the weight equivalent mean if the density is constant.

So each technique is liable to generate a different mean diameter as well as measuring different properties of our particle. No wonder people get confused! There are also an infinite number of "right" answers. Imagine 3 spheres with diameters 1, 2, 3 units:

$$X_{nl} = D[1,0] = \frac{1 + 2 + 3}{3} = 2.00$$

$$X_{ns} = D[2,0] = \sqrt{\frac{1 + 4 + 9}{3}} = 2.16$$

$$X_{nv} = D[3,0] = \sqrt[3]{\frac{1 + 8 + 27}{3}} = 2.29$$

$$X_{ls} = D[2,1] = \frac{1 + 4 + 9}{1 + 2 + 3} = 2.33$$

$$X_{lv} = D[3,1] = \sqrt{\frac{1 + 8 + 27}{1 + 2 + 3}} = 2.45$$

$$X_{sv} = D[3,2] = \frac{1 + 8 + 27}{1 + 4 + 9} = 2.57$$

$$X_{vm} = D[4,3] = \frac{1 + 16 + 81}{1 + 8 + 27} = 2.72$$

$$= X_{wm}$$

Number and volume distributions

Size (cm)	Number of Objects	% by Number	% by Mass
10-1000	7000	0.2	99.96
1-10	17500	0.5	0.03
0.1-1.0	3500000	99.3	0.01
Total	3524500	100.00	100.00

The above example is adapted from an article in New Scientist (13 October 1991). There are a large number of man-made objects orbiting the earth in space and scientists track them regularly. Scientists have also classified them in groups on the basis of their size.

If we examine the third column above we would conclude (correctly) that 99.3% of all particles are incredibly small. This is evaluating the data on a NUMBER basis. However, if we examine the fourth column we would conclude (correctly) that virtually all the objects are between 10 – 1000cms. This is where all the MASS of the object is. Note that the NUMBER and MASS distribution are very different and we would draw different conclusions depending on

which distributions we use.

Again neither distribution is incorrect. The data are just being examined in different ways. If we were making a space suit, for example, we could say that it is easy to avoid the 7000 large objects and this takes care of 99.96% of all cases. However, what is more important with a space suit is the protection against small particles which are 99.3% by number!

If we take a calculator and calculate the means of the above distributions we find that the number mean is about 1.6cm and the mass mean about 500cm – again very different.

Interconversion between number, length and volume/mass means.

If we are measuring our particles on an electron microscope we know, from an earlier section (Different techniques give different means.) that we are calculating the $D[1,0]$ or the number-length mean size. If what we really require is the mass or volume mean size we have to convert our number mean to a mass mean. Mathematically, this is easily feasible, but let us examine the consequences of such a conversion.

Imagine that our electron measurement technique is subject to an error of $\pm 3\%$ on the mean size. When we convert the number mean size to a mass mean size then as the mass mean is a cubic function of the diameter then our errors will be cubed or $\pm 27\%$ variation on the final result.

However, if we are calculating the mass or volume distribution as we do with laser diffraction then the situation is different. For a stable sample measured under recirculating conditions in liquid suspension, we should be able to generate a volume mean reproducibility of $\pm 0.5\%$. If now we convert this volume mean to a number mean the error or the number mean is the cube root of 0.5% or less than 1.0% !

In practice this means that if we are using an electron microscope and what we really want is a volume or mass distribution, the effect of ignoring or missing one 10μ particle is the same as ignoring or missing one thousand 1μ particles. Thus we must be aware of the great dangers of interconversion.



On the Malvern Sizers both the DOSTM and WindowsTM software will calculate other derived diameters but we must be very careful of how we interpret these derived diameters. Different means can be converted to each other by means of the following equations (Hatch-Choate transformation) (Ref.7):

$$\ln D_{4,3} = 1/\ln X_v + 0.5/\ln^2 \sigma$$

$$\ln D_{4,2} = 2/\ln X_v$$

$$\ln D_{4,1} = 3/\ln X_v - 1.5/\ln^2 \sigma$$

$$\ln D_{4,0} = 4/\ln X_v - 4/\ln^2 \sigma$$

$$\ln D_{3,2} = 1/\ln X_v - 0.5/\ln^2 \sigma$$

$$\ln D_{3,1} = 2/\ln X_v - 2/\ln^2 \sigma$$

$$\ln D_{3,0} = 3/\ln X_v - 4.5/\ln^2 \sigma$$

$$\ln D_{2,0} = 2/\ln X_v - 4/\ln^2 \sigma$$

$$\ln D_{2,1} = 1/\ln X_v - 1.5/\ln^2 \sigma$$

$$\ln D_{1,0} = 3/\ln X_v - 2.5/\ln^2 \sigma$$

Measured and derived diameters.

We have seen that the Malvern laser diffraction technique generates a volume distribution for the analysed light energy data. (Note that with Fraunhofer analysis, the projected area distribution is assumed). This volume distribution can be converted to any number or length diameter as shown above.

However, in any analysis technique, we must be aware of the consequences of such a conversion (see previous section "Interconversion between number, length and volume/mass means.") and also which mean diameter is actually measured by the equipment and which diameters are really calculated or derived from that first measured diameter.

Other techniques will generate other diameters from some measured

diameters. For example, a microscope will measure the $D[1,0]$ and will/may derive other diameters from this.

We can place more faith in the measured diameter than we can on the derived diameters. In fact, in some instances it can be very dangerous to rely on the derived property. For example, the Malvern analysis table gives us a specific surface area in m^2/cc or m^2/gm . We must not take this value too literally – in fact, if what we really want is the specific surface area of our material we really should use a surface area specific technique e.g. B.E.T. or mercury porosimetry.

Which number do we use?

Remembering that each different technique measures a different property (or size) of our particle and that we may use the data in a number of different ways to get a different mean result ($D[4,3]$, $D[3,2]$ etc.), then what number should we use?

Let's take a simple example of two spheres of diameters 1 and 10 units. Imagine that we are making gold. If we calculate the simple number mean diameter this will give us:

$$D[1,0] = \frac{1+10}{2} = 5.50$$

So we would assume that the average size of the particles in the system is 5.50 units. However, we must remember that if we are making gold we are interested in the weight of our material.

For example, if we have a process stream we are not interested that there are 3.5 million particles in it, we are more interested that there is 1kg or 2kg of gold.

Remembering that the mass mean is a cubic function of diameter, we would see that the sphere of diameter 1 unit has a mass of 1 unit and the sphere of diameter 10 units has a mass of $10^3 = 1000$ units.

That is, the larger sphere makes up 1000/1001 parts of the total mass of the system. If we are making gold then we can throw away the sphere of 1 unit because we will be losing less than 0.1% of the total mass of the system. So the number mean does not accurately reflect where the mass

of the system lies. This is where the $D[4,3]$ is much more useful.

In our two sphere example the mass or volume moment mean would be calculated as follows:

$$D[4,3] = \frac{1^4 + 10^4}{1^3 + 10^3} = 9.991$$

This value shows us more where the mass of the system lies and is of more value to chemical process engineers.

However, let us imagine that we are in a clean room making wafers of silicon or gallium arsenide. Here, if one particle lands on our wafer it will tend to produce a defect. In this instance the number or concentration of the particles is very important because 1 particle = 1 defect. We would want to use a technique that directly measures the number of particles or gives us the concentration of particles. In essence this is the difference between particle counting and particle sizing. With counting we will record each particle and count it – the size is less important and we may only require a limited number of size classes (say 8). With sizing the absolute number of particles is less relevant than the sizes or the size distribution of the particles and we may require more size bands.

For a metered dose inhaler for asthma sufferers then both the concentration of drug and its particle size distribution is important.

Mean, Median and Mode – basic statistics

It is important to define these three terms as they are so often misused in both statistics and particle size analysis:

MEAN

This is some arithmetic average of the data. There are a number of means that can be calculated for particles (see section $D[4,3]$ etc.).

MEDIAN

This is the value of the particle size which divides the population exactly into two equal halves i.e. there is 50% of the distribution above this value and 50% below.



MODE

This is the most common value of the frequency distribution i.e. the highest point of the frequency curve.

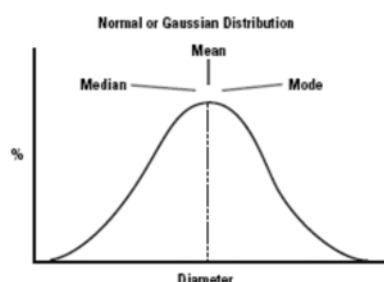


Figure 4

Imagine that our distribution is a Normal or Gaussian distribution. The mean, median and mode will lie in exactly the same positions. See Figure 4.

However, imagine that our distribution is bimodal as shown in Figure 5.

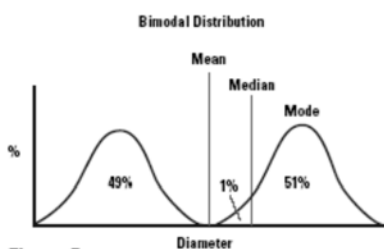


Figure 5

The mean diameter will be almost exactly between the two distributions as shown. Note there are no particles which are this mean size! The median diameter will lie 1% into the higher of the two distributions because this is the point which divides the distribution exactly into two. The mode will lie at the top of the higher curve because this is the most common value of the diameter (only just!).

This example illustrates that there is no reason which the mean, median and mode should be identical or even similar, it depends on the symmetry of the distribution.

Note that in the Malvern analysis table:

- $D[4,3]$ is the volume or mass moment mean or the De Broucker mean.
- $D[\nu,0.5]$ is the volume (ν) median diameter sometimes shown as D_{50} or $D_{0.5}$.

- $D[3,2]$ is the surface area moment mean or the Sauter Mean Diameter (SMD).

Methods of measurement

From our earlier sections, we have seen that each measurement technique produces a different answer because it is measuring a different dimension of our particle. We will now discuss some of the relative advantages and disadvantages of the main different methods employed.

Sieves

This is an extremely old technique but has the advantage that it is cheap and is readily usable for large particles such as are found in mining. Terence Allen (Ref. 2) discussed the difficulties of reproducible sieving but the main disadvantages to many users are the following:

- Not possible to measure sprays or emulsions
- Measurement for dry powders under 400# (38 μ) very difficult. Wet sieving is said to solve this problem but results from this technique give very poor reproducibility and are difficult to carry out.
- Cohesive and agglomerated materials e.g. clays are difficult to measure.
- Materials such as 0.3 μ TiO₂ are simply impossible to measure and resolve on a sieve. The method is not inherently high resolution.
- The longer the measurement, the smaller the answer as particles orientate themselves to fall through the sieve. This means that measurement times and operating methods (e.g. tapping) need to be rigidly standardised.
- A true weight distribution is not produced. Rather the method relies on measuring the second smallest dimension of the particle. This can give some strange results with rod-like materials e.g. paracetamol in the pharmaceutical industry.
- Tolerance. It is instructive to examine a table of ASTM or BS sieve sizes and see the permitted tolerances on average and maximum variation. The reader is invited to do this.

Sedimentation

This has been the traditional method of measurement in the paint and ceramics industry and gives seductively low answers! The applicable range is 2-50 microns (Ref. 1 & 2) despite what the manufacturers may claim.

The principle of measurement is based on the Stokes' Law equation:

$$\text{Terminal Velocity, } U_s = \frac{(p_s - p_f) D^2 g}{18\eta}$$

Equipment can be as simple as the Andreason pipette or more complicated involving the use of centrifuges or X-rays.

Examination of this equation will indicate one or two potential pitfalls. The density of the material is needed, hence the method is no good for emulsions where the material does not settle, or very dense materials which settle quickly. The end result is a Stokes diameter (D_{ST}) which is not the same as a weight diameter, $D[4,3]$, and is simply a comparison of the particle's settling rate to a sphere settling at the same rate. The viscosity term in the denominator indicates that we will need to control temperatures very accurately – a 1°C change in temperature will produce a 2% change in viscosity.

With the equation it is relatively easy to calculate settling times. It can be shown that a one micron particle of SiO₂ ($\rho = 2.5$) will take 3.5 hours to settle 1cm under gravity in water at 20°C. Measurements are therefore extremely slow and repeat measurements tedious. Hence the move to increase g and attempt to remedy the situation.

The disadvantages of increasing ' g ' are discussed in (Ref. 3). More specific criticisms of the sedimentation technique are to be found in (Ref. 2).

Stokes' Law is only valid for spheres which possess the unique feature of being the most compact shape for the volume or surface area they possess. Hence more irregularly shaped 'normal' particles will possess more surface area than the sphere and will therefore fall more slowly because of the increased drag than their equivalent spherical diameters.



Comparison of Brownian movement displacement and gravitational settling displacement

Displacement in 1.0 second (μm)						
Particle diameter (μm)	In air at 70°F (1atm)		In water at 70°F		In water at 70°F	
	Due to Brownian movement*	Due to Gravitational settling+	Due to Brownian movement*	Due to Gravitational settling+	Y	$k = \frac{100Y}{\%}$
0.10	29.4	1.73	2.36	0.005	31.1	96.9
0.25	14.2	6.3	1.49	0.0346	3.15	75.9
0.50	8.92	19.9	1.052	0.1384	0.556	35.7
1.0	5.91	69.6	0.745	0.554	0.0983	5.0
2.5	3.58	400	0.334	13.84	0.00995	1.0
10.0	1.75	1550	0.236	55.4	0.00031	0.03

*Mean displacement given by equation (7.20)

+Distance settled by a sphere of density 2000kgm³, including Cunningham's correction.

Y is defined in equation (7.23) (Taken from Reference 2 p 259)

For objects like kaolins which are disc-shaped this effect is even more accentuated and large deviations from reality are to be expected.

Furthermore, with small particles there are two competing processes – gravitational settling and Brownian motion. Stokes' Law only applies to gravitational settling. The table at the top of this page shows a comparison between the two competing processes. It will be seen that very large errors (approx. 20%) will result if sedimentation is used for particles under 2μm in size and the errors will be in excess of 100% for 0.5μm particles.

The sedimentation technique gives an answer smaller than reality and this is why some manufacturers deceive themselves. In summary the main disadvantages of the technique for pigment users are the following:

- Speed of measurement. Average times are 25 minutes to 1 hour for measurement making repeat analyses difficult and increasing the chances for reagglomeration.
- Accurate temperature control. Needed to prevent temperature gradients and viscosity changes.
- Inability to handle mixtures of differing densities – many pigments are a mixture of colouring matter and extender/filler.
- Use of X-rays. Some systems use X-rays and, in theory, personnel should be monitored.
- Limited range. Below 2μm, Brownian motion predominates

and the system is inaccurate.

Above 50μm, settling is turbulent and Stokes' Law again is not applicable.

Figure 6 shows the expected differences between a sedimentation and laser diffraction results.

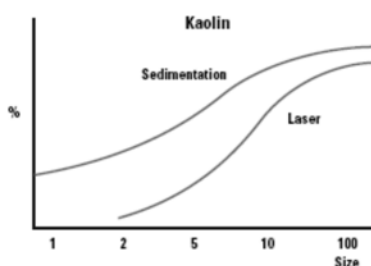


Figure 6

Electrozone sensing (Coulter Counter)

This technique was developed in the mid 1950's for sizing blood cells which are virtually monomodal suspension in a dilute electrolyte.

The principle of operation is very simple. A glass vessel has a hole or orifice in it. Dilute suspension is made to flow through this orifice and a voltage applied across it. As particles flow through the orifice the capacitance alters and this is indicated by a voltage pulse or spike. In older instruments the peak height was measured and related to a peak height of a standard latex. Hence the method is not an absolute one but is of a comparative nature. Problems of particle orientation through the beam

can be corrected for by measuring the area under the peak rather than the peak height. For blood cells the technique is unsurpassed and the method is capable of giving both a number count and volume distribution. For real, industrial materials such as pigments there are a number of fundamental drawbacks:

- Difficult to measure emulsions. (Sprays not possible!) Dry powders need to be suspended in a medium so cannot be measured directly.
- Must measure in an electrolyte. For organic based materials this is difficult as it is not possible to measure in xylene, butanol and other poorly conducting solutions.
- The method requires calibration standards which are expensive and change their size in distilled water and electrolyte (Ref. 2).
- For materials of relatively wide particle size distribution the method is slow as orifices have to be changed and there is a danger of blocking the smaller orifices.
- The bottom limit of the method is set by the smallest orifice available and it is not easy to measure below 2μm or so. Certainly it is not possible to measure TiO₂ at 0.2μm.
- Porous particles give significant errors as the "envelope" of the particle is measured.



- Dense materials or large materials are difficult to force through the orifice as they sediment before this stage.

So, in summary this technique is excellent for blood cells but of a more dubious nature for many industrial materials.

Microscopy

This is an excellent technique as it allows one to directly look at the particles in question. So the shape of the particles can be seen and it can also be used to judge whether good dispersion has been achieved or whether agglomeration is present in the system. The method is relatively cheap and for some microscope systems it is possible to use image analysis to obtain lists of numbers (usually to 6 or 8 places of decimals, well beyond the resolution of the technique!).

It is interesting to note that 1g of 10µm particles (density 2.5) contains 760×10^6 particles – all these can never be examined individually by microscopy.

However, it is not suitable as a quality or production control technique beyond a simple judgement of the type indicated above. Relatively few particles are examined and there is the real danger of unrepresentative sampling. Furthermore, if a weight distribution is measured the errors are magnified – missing or ignoring one 10µm particle has the same effect as ignoring one thousand 1µm particles.

Electron microscopy has elaborate sample preparation and is slow. With manual microscopy few particles are examined (maybe 2000 in a day with a good operator) and there is rapid operator fatigue. Again there is the problem of "which dimension do we measure?" Hence there can be large operator to operator variability on the same sample. In combination with diffraction microscopy becomes

a very valuable aid to the characterization of particles.

Laser diffraction

This is more correctly called Low Angle Laser Light Scattering (LALLS). This method has become the preferred standard in many industries for characterization and quality control. The applicable range according to ISO13320 is 0.1 – 3000µm. Instrumentation has been developed in this field over the last twenty years or so. The method relies on the fact that diffraction angle is inversely proportional to particle size.

Instruments consist of:

- A laser as a source of coherent intense light of fixed wavelength. He-Ne gas lasers ($\lambda=0.63\mu\text{m}$) are the most common as they offer the best stability (especially with respect to temperature) and better signal to noise than the higher wavelength laser diodes. It is to be expected when smaller laser diodes can reach 600nm and below and become more reliable that these will begin to replace the bulkier gas lasers.
- A suitable detector. Usually this is a slice of photosensitive silicon with a number of discrete detectors. It can be shown that there is an optimum number of detectors (16–32) – increased numbers do not mean increased resolution. For the photon correlation spectroscopy technique (PCS) used in the range 1nm – 1µm approximately, the intensity of light scattered is so low that a photomultiplier tube, together with a signal correlator is needed to make sense of the information.
- Some means of passing the sample through the laser beam. In practice it is possible to measure aerosol sprays directly by spraying them through the beam. This makes a traditionally difficult measurement extremely simple. A dry powder can be blown

through the beam by means of pressure and sucked into a vacuum cleaner to prevent dust being sprayed into the environment. Particles in suspension can be measured by recirculating the sample in front of the laser beam.

Older instruments and some existing instruments rely only on the Fraunhofer approximation which assumes:

- Particle is much larger than the wavelength of light employed (ISO13320 defines this as being greater than 40λ i.e. 25µm when a He-Ne laser is used).
- All sizes of particle scatter with equal efficiencies.
- Particle is opaque and transmits no light.

These assumptions are never correct for many materials and for small material they can give rise to errors approaching 30% especially when the relative refractive index of the material and medium is close to unity. When the particle size approaches the wavelength of light the scattering becomes a complex function with maxima and minima present. The latest instruments (e.g. Mastersizer 2000, Malvern Instruments) use the full Mie theory which completely solves the equations for interaction of light with matter. This allows completely accurate results over a large size range (0.02 – 2000µm typically). The Mie theory assumes the volume of the particle as opposed to Fraunhofer which is a projected area prediction.

The "penalty" for this complete accuracy is that the refractive indices for the material and medium need to be known and the absorption part of the refractive index known or guessed. However, for the majority of users this will present no problems as these values are either generally known or can be measured.



Laser diffraction gives the end-user the following advantages:

- The method is an absolute one set in fundamental scientific principles. Hence there is no need to calibrate an instrument against a standard – in fact there is no real way to calibrate a laser diffraction instrument. Equipment can be validated, to confirm that it is performing to certain traceable standards.
- A wide dynamic range. The best laser diffraction equipment allows the user to measure in the range from say 0.1 to 2000 microns. Smaller samples (1nm – 1µm) can be measured with the photon correlation spectroscopy technique as long as the material is in suspension and does not sediment.
- Flexibility. For example it is possible to measure the output from a spray nozzle in a paint booth. This has been used by nozzle designers, to optimise the viscosity, ΔP and hole size and layout, in order to get correct droplet size. This has found extensive application in the agricultural and pharmaceutical industries. For further information the reader is referred to References 4,5 and 6. There is now an ASTM standard for sprays using laser diffraction.
- Dry powders can be measured directly, although this may result in poorer dispersion than using a liquid dispersing medium. However, in conjunction with a suspension analysis it can be valuable in assessing the amount of agglomerated material in the dry state.
- Liquid suspensions and emulsions can be measured in a recirculating cell and this gives high reproducibility and also allows dispersing agents (e.g. 0.1% Calgon, sodium hexametaphosphate solution for TiO_2) and surfactants to be employed to ascertain the primary particle size. If possible the preferred method would be to measure in liquid suspension (aqueous or organics) for the reasons discussed above.
- The entire sample is measured. Although samples are small (4-10g for dry powders, 1-2g for suspensions typically) and a representative sample must be obtained, all the sample passes through the laser beam and diffraction is obtained from all the particles.
- The method is non-destructive and non-intrusive. Hence samples can be recovered if they are valuable.
- A volume distribution is generated directly which is equal to the weight distribution if the density is constant. This is the preferred distribution for chemical engineers.
- The method is rapid producing an answer in less than one minute. This means rapid feedback to operating plants and repeat analyses are made very easily.
- Highly repeatable technique. This means that the results can be relied on and the plant manager knows that his product has genuinely changed and that the instrument is not "drifting".
- High resolution. Up to 100 size classes within the range of the system can be calculated on the Malvern Mastersizer.

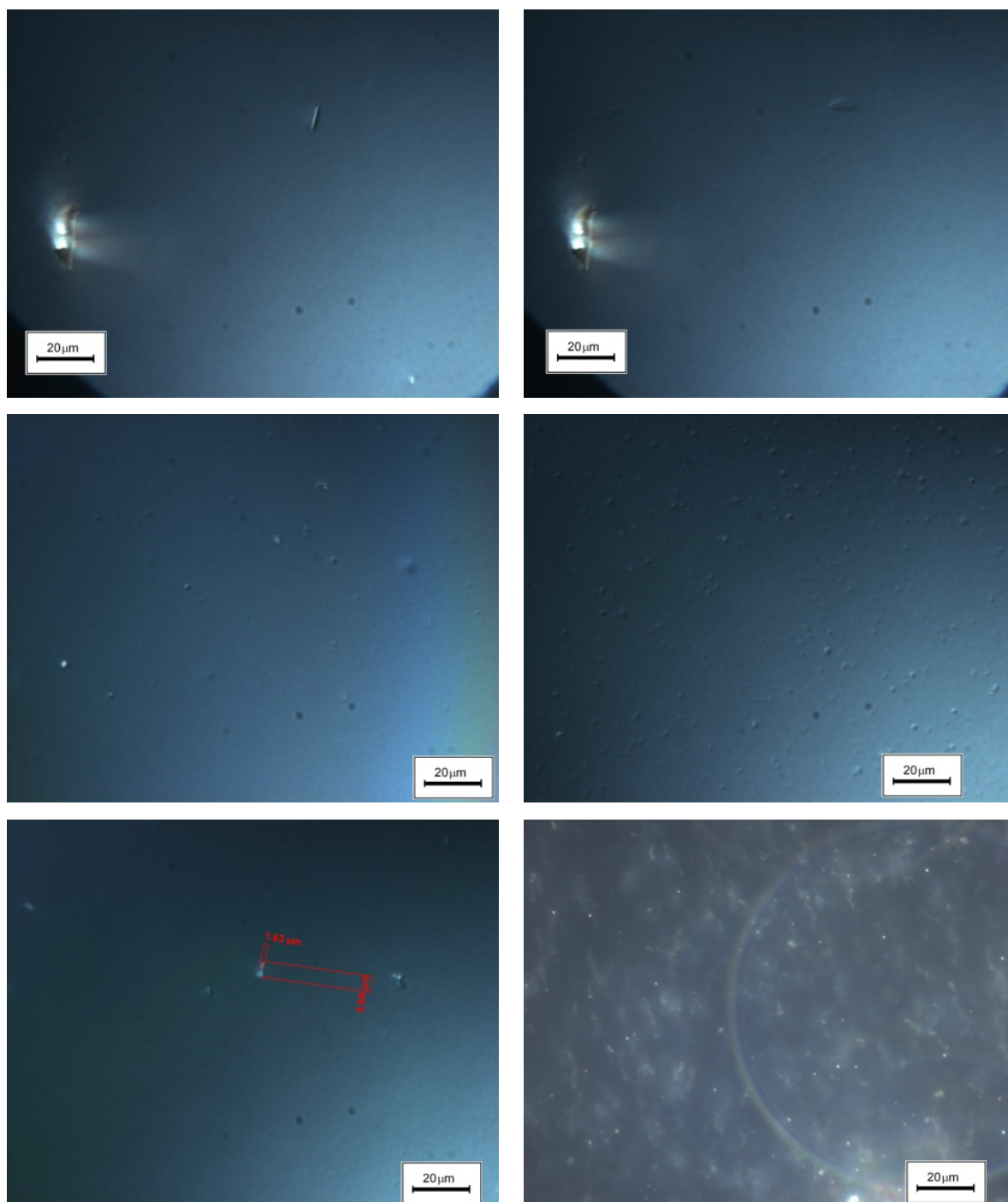
References

1. *Paint and Surface coatings – theory and practice*; Ed. R. Lambourne Ellis Horwood Ltd. 1993. ISBN 0-13-030974-5PGk
2. *Particle Size Measurement*; T. Allen, Chapman & Hall. 4th Edition, 1992. ISBN 04123570
3. G.J.J. Beckers, H.J. Veringa; *Powder Technology* 60 (1989) p245-248.
4. J. Ranucci *Pharmaceutical Technology*; October 1992 pp 108-114.
5. G. Hind; *Manufacturing Chemist* August 1990 pp 28-30.
6. M.W. Wedd; *ILASS-Europe 8th Annual conference, Koninklijke/Shell Laboratorium, Amsterdam 30 September –2nd October, 1992.*
7. T. Hatch & S.P. Choate. *J. Franklin Inst.* 207 pp 369-387 (1929).



E. Microscopia òptica dels agregats de PEO-b-PCL obtinguts per diàlisi

A continuació es mostren totes les imatges de microscopia òptica que no s'han pogut introduir a la memòria.



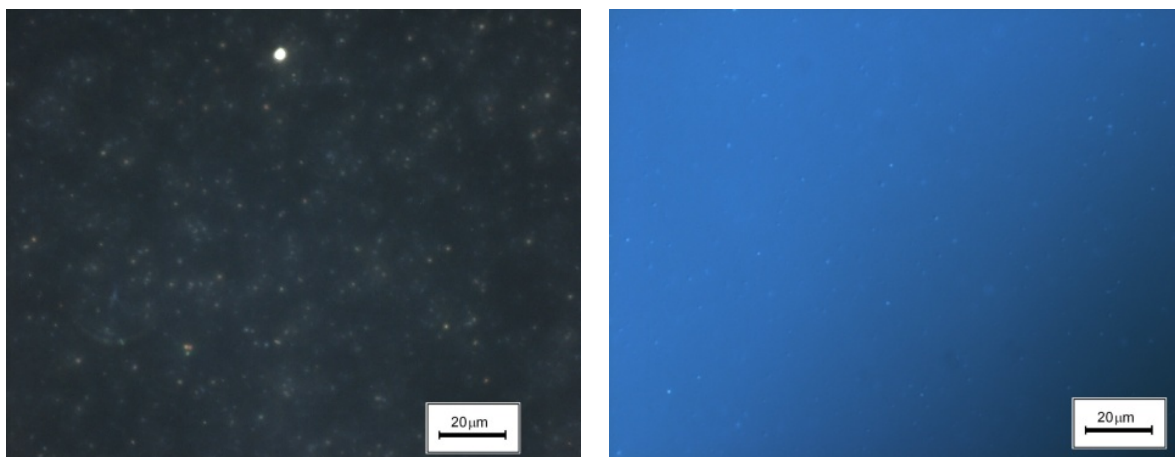


Figura E.1 Imatges de microscopia òptica de les dissolucions d'agregats. (a) $\text{PEO}_{2000}\text{-}b\text{-PCL}_{4000}$ (IC); (b) $\text{PEO}_{2000}\text{-}b\text{-PCL}_{4000}$ (IC); (c) $\text{PEO}_{2000}\text{-}b\text{-PCL}_{6000}$ (IC); (d) $\text{PEO}_{2000}\text{-}b\text{-PCL}_{8000}$ (IC); (e) $\text{PEO}_{5000}\text{-}b\text{-PCL}_{10000}$ (IC); (f) $\text{PEO}_{5000}\text{-}b\text{-PCL}_{15000}$ (DF) ; (g) $\text{PEO}_{5000}\text{-}b\text{-PCL}_{15000}$ (DF) ; (h) $\text{PEO}_{5000}\text{-}b\text{-PCL}_{20000}$ (IC)

A la carpeta VIDEOS del CD s'hi troben els següents vídeos:

2-6ICx50

2-8ICx50

5-10ICx50

5-15DFx50_1

5-15DFx50_2

5-20ICx50

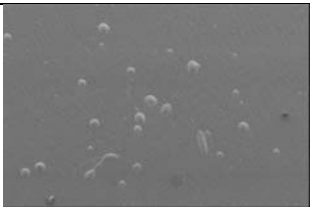
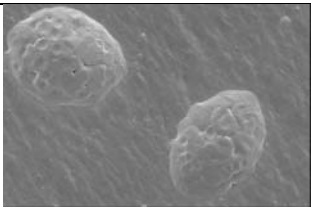
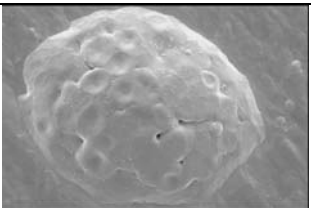
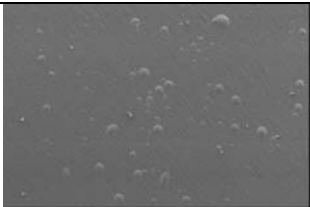
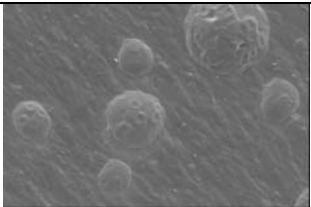
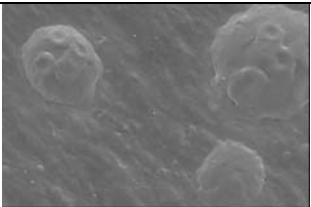
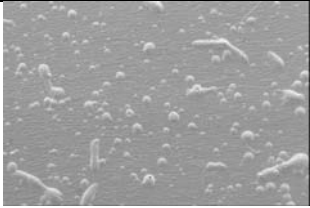
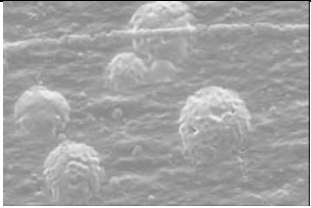
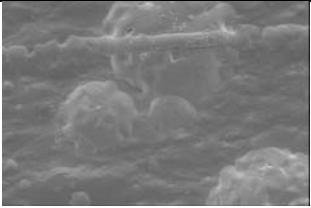
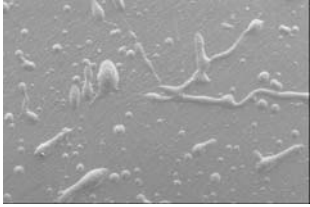
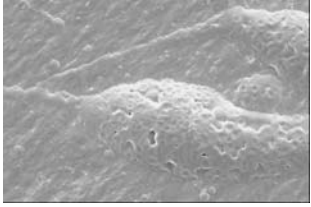
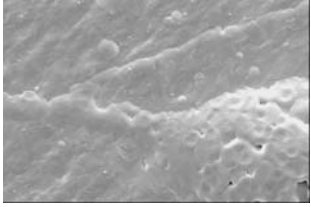
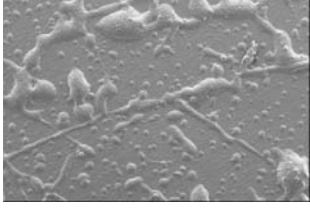
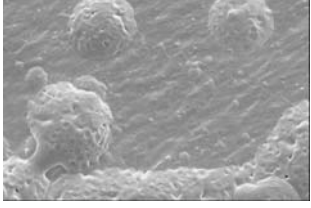
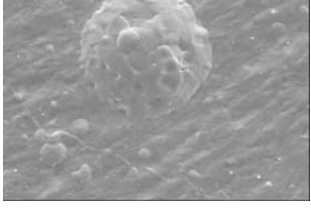
x-y	identificació del copolímer
IC	contrast per interferències
DF	camp fosc
x50	augments



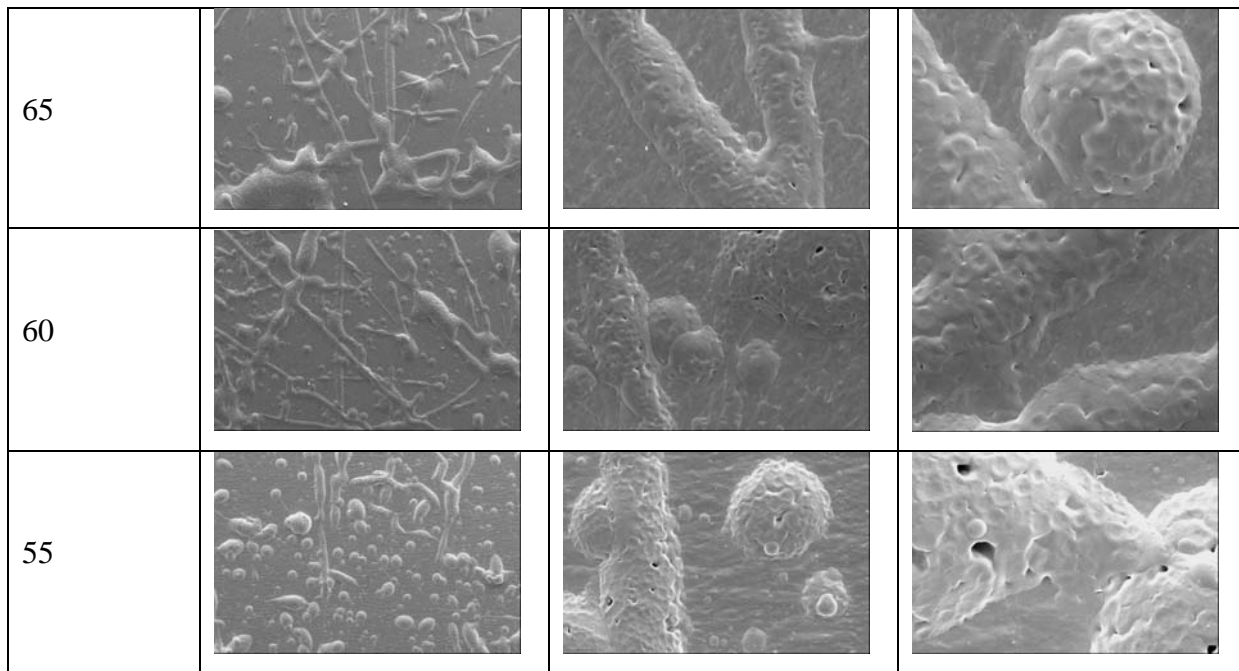
F. Imatges de SEM dels copolímers fosos de PEO-b-PCL per *electrospinning*

F.1 Estudi del paràmetre temperatura

Taula F.1 Imatges de SEM per a l'estudi de l'efecte de la temperatura en el copolímer PEO₂₀₀₀-*b*-PCL₄₀₀₀

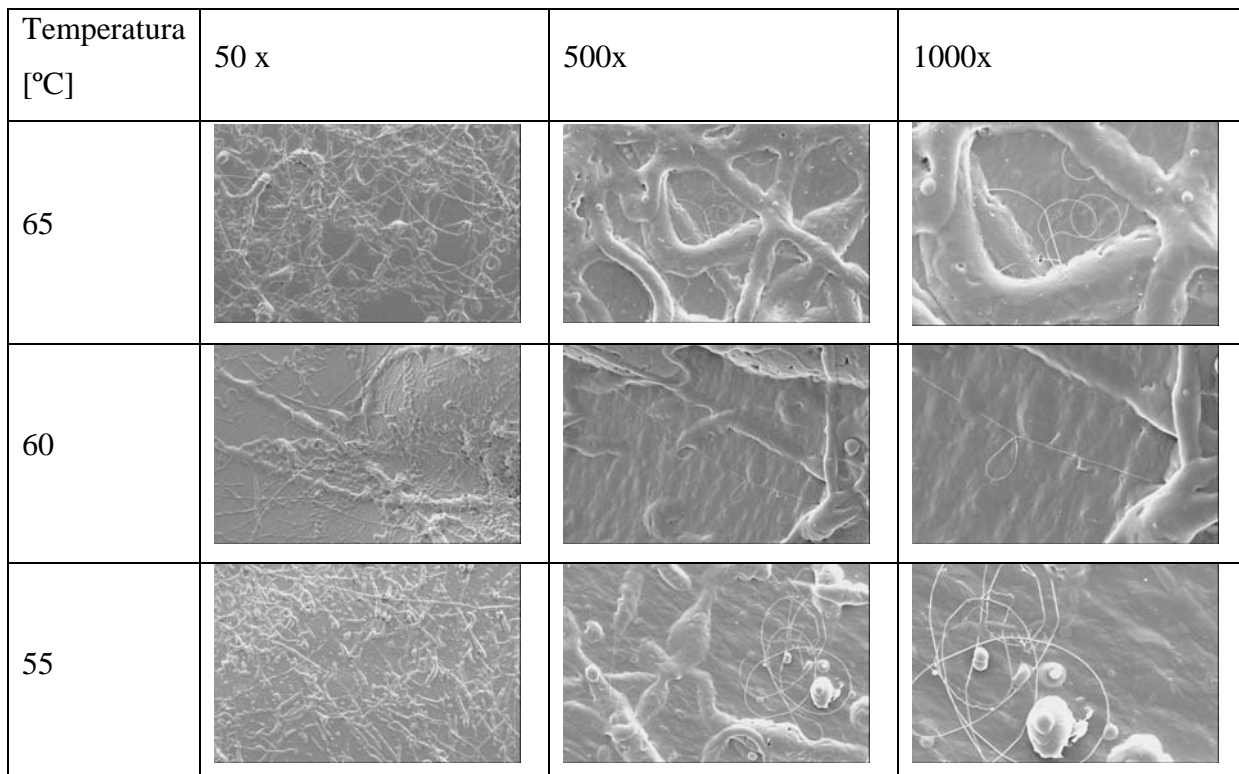
Temperatura [°C]	50 x	500x	1000x
90			
85			
80			
75			
70			





Per al copolímer $\text{PEO}_{2000}\text{-}b\text{-PCL}_{6000}$ s'han descartat les mostres entre 70 – 90 °C perquè no hi havia més que fibres foses en peces molt grans.

Taula F.2 Imatges de SEM per a l'estudi de l'efecte de la temperatura en el copolímer $\text{PEO}_{2000}\text{-}b\text{-PCL}_{6000}$



Per al copolímer PEO₂₀₀₀-*b*-PCL₄₀₀₀ no s'han guardat imatges de SEM entre 85 – 70 °C degut a la manca de fibres seques i a la presència exclusiva de fibres foses de mides molt grans. La imatge de la Figura F.1 se'n pot considerar representativa.

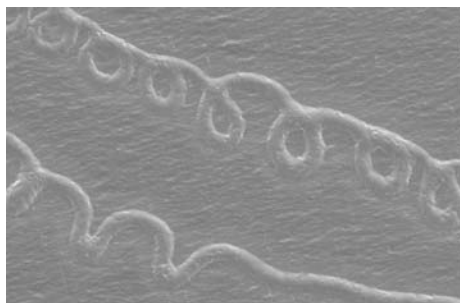


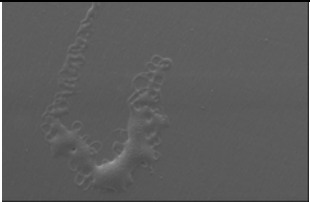
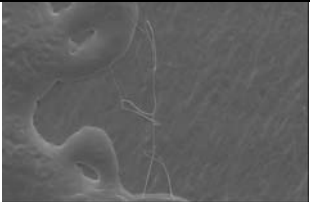

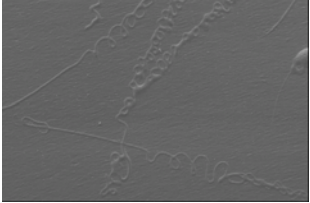
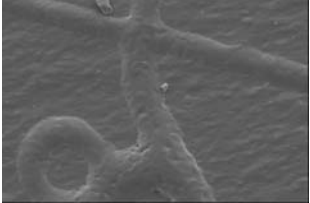

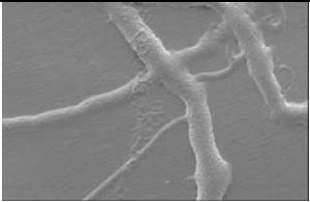
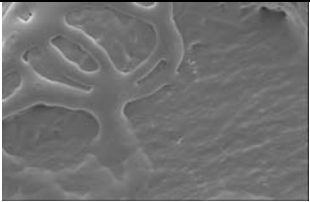
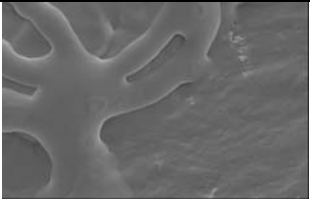



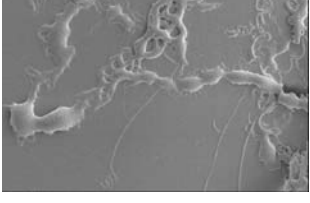
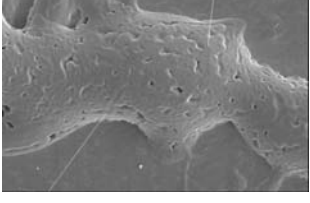
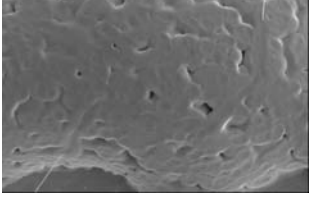
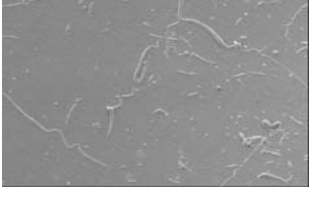


Figura F.1 Imatge de SEM d'una mostra del copolímer PEO₂₀₀₀-*b*-PCL₈₀₀₀ representativa per a les mostres en el rang de temperatura 70-85 °C.

Taula F.3 Imatges de SEM per a l'estudi de l'efecte de la temperatura en el copolímer PEO₂₀₀₀-*b*-PCL₈₀₀₀

Temperatura [°C]	50 x	500x	1000x
90			
65			
60			
55			



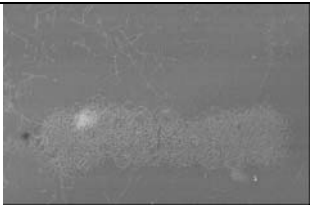
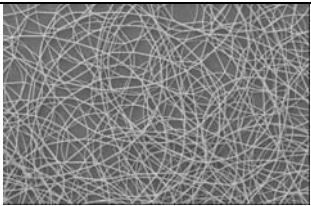
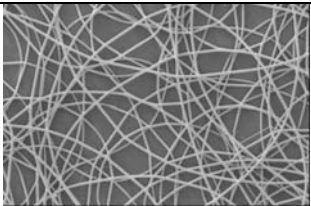
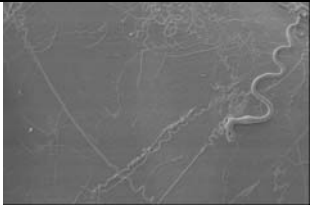
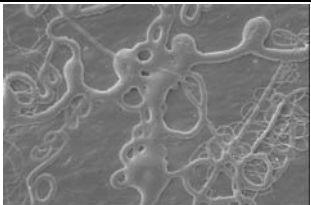

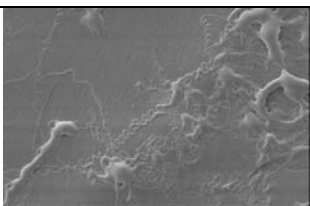
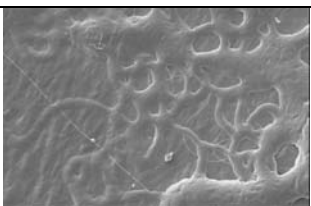
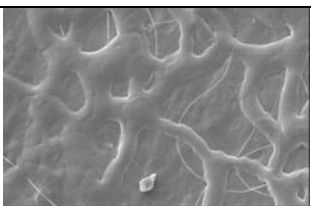
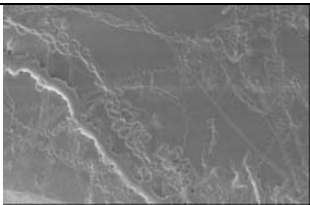
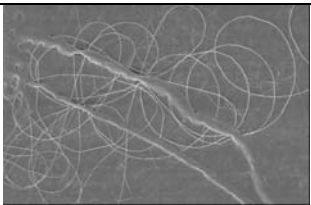
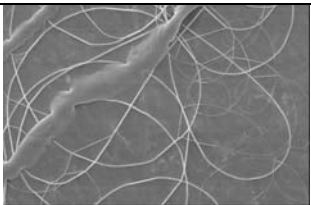
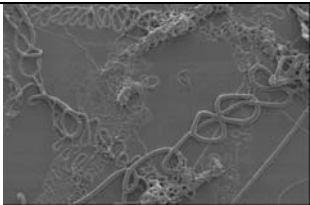
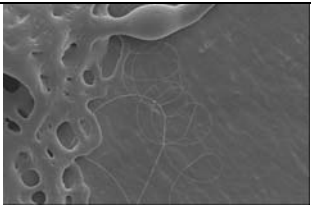
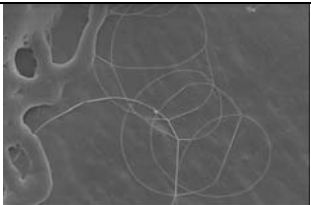
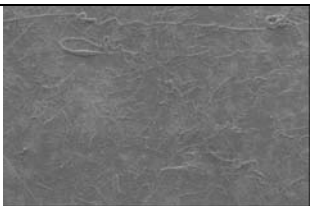
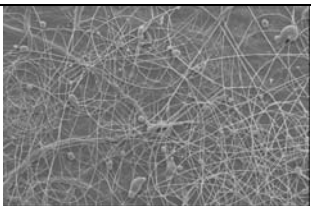
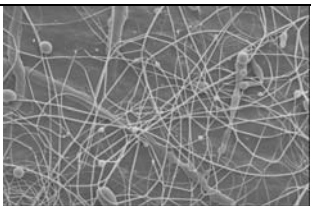
Taula F.4 Imatges de SEM per a l'estudi de l'efecte de la temperatura en el copolímer PEO₅₀₀₀-*b*-PCL₁₀₀₀₀

Temperatura [°C]	50 x	500x	1000x
90			
85			
80			
75			
70			
65			



Tal i com s'ha explicat a la memòria, els dos copolímers que presenten zones no aïllades de fibres seques són el PEO₅₀₀₀-*b*-PCL₁₅₀₀₀ i PEO₅₀₀₀-*b*-PCL₂₀₀₀₀. En el cas del PEO₅₀₀₀-*b*-PCL₁₅₀₀₀ també s'ha observat una àrea d'elevada concentració de fibres seques (90°C).

Taula F.5 Imatges de SEM per a l'estudi de l'efecte de la temperatura en el copolímer PEO₅₀₀₀-*b*-PCL₁₅₀₀₀

Temperatura [°C]	50 x	500x	1000x
90			
85			
80			
75			
70			
65			



Taula F.6 Imatges de SEM per a l'estudi de l'efecte de la temperatura en el copolímer PEO₅₀₀₀-*b*-PCL₂₀₀₀₀

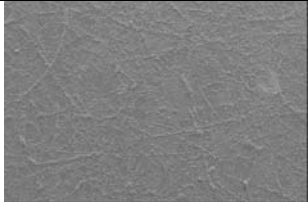
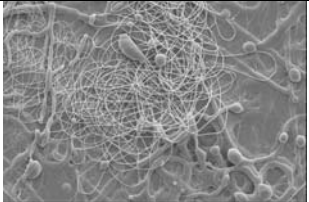
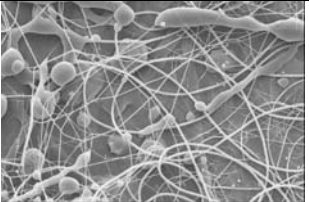
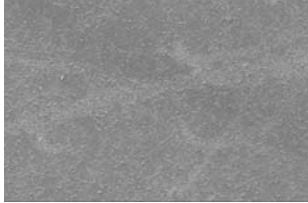
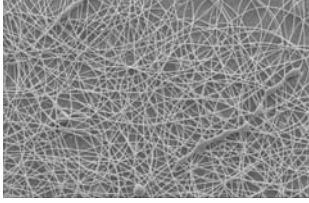
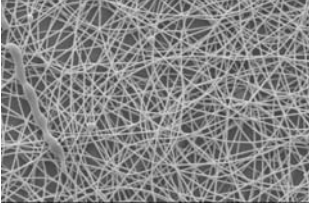
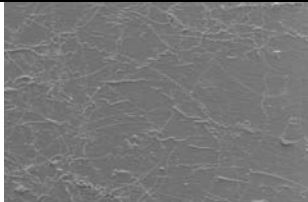
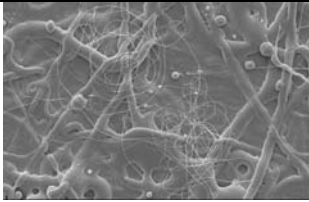
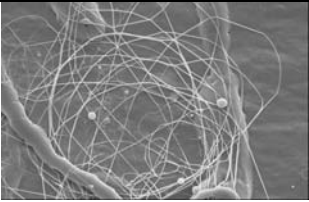
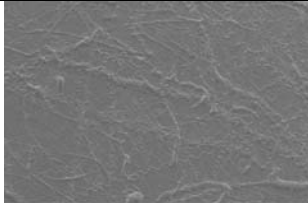


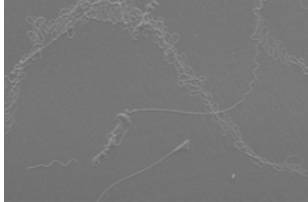
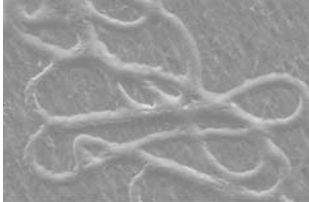
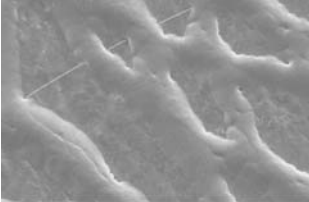
Temperatura [°C]	50 x	500x	1000x
90			
85			
80			
75			
70			
65			



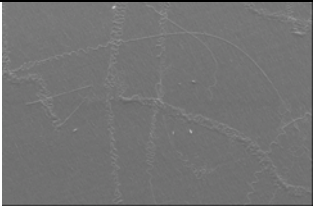
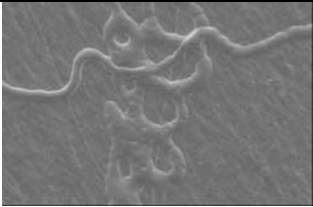
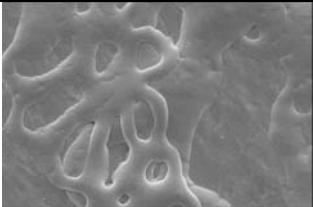




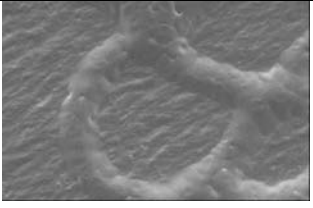
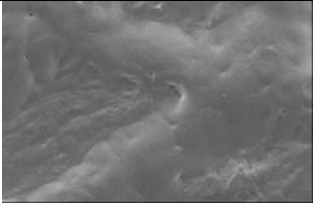
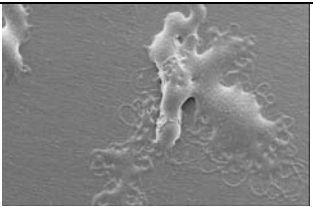
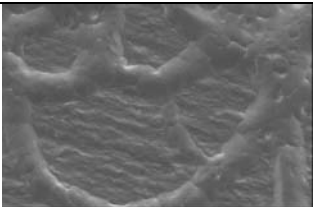
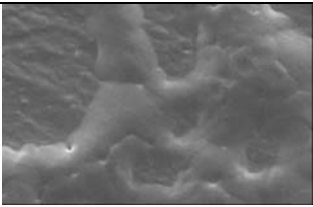
F.2 Estudi del paràmetre cabal

El cabal s'ha estudiat per al PEO₅₀₀₀-*b*-PCL₁₅₀₀₀ i PEO₅₀₀₀-*b*-PCL₂₀₀₀₀ entre 0.01 ml/h i 7.5 ml/h però a 0.01 ml/h no s'han pogut produir les mostres per manca de polímer. Es destaca l'augment notable del diàmetre de les fibres, especialment als cabals més grans, així com la presència de fibres seques només en la primera meitat de cabals. Totes les mostres per a l'estudi del cabal s'han realitzat a 85 °C.

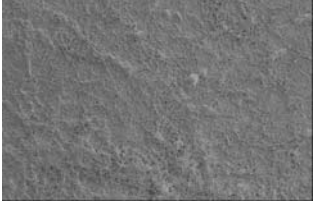
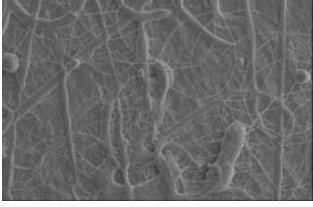
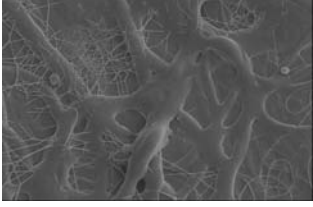
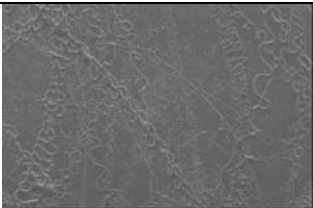
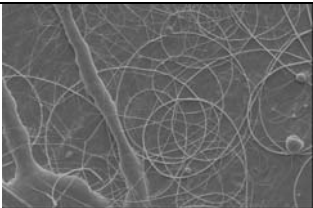
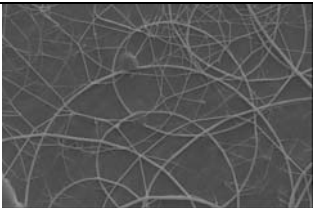
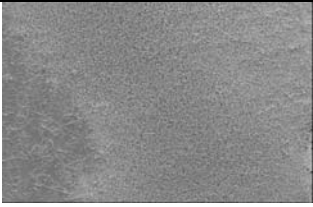
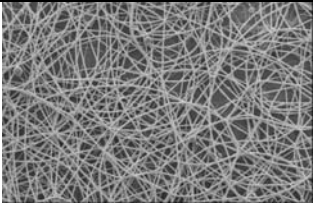
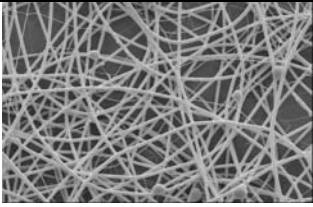
Taula F.7 Imatges de SEM per a l'estudi de l'efecte del cabal en el copolímer PEO₅₀₀₀-*b*-PCL₁₅₀₀₀

Cabal [ml/h]	50 x	500x	1000x
0.02			
0.05			
0.1			
0.2			
0.5			

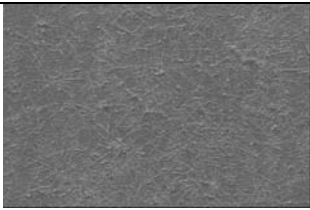
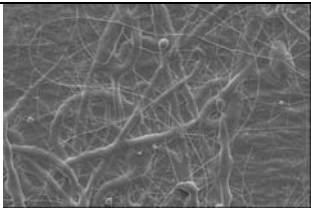
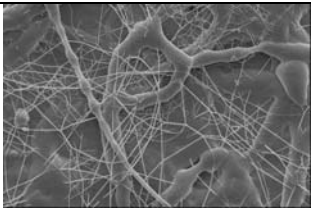
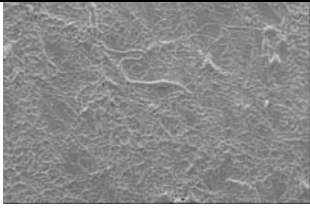
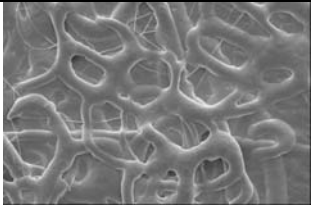
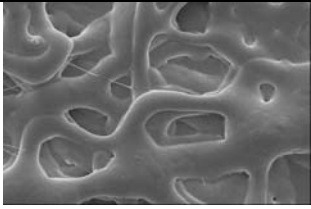
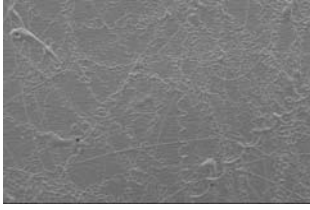
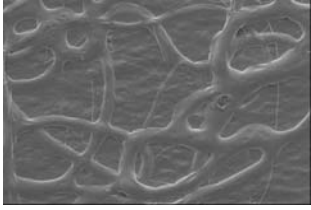
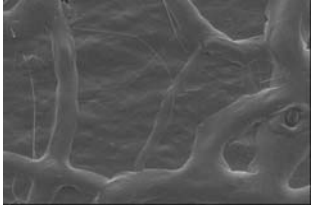
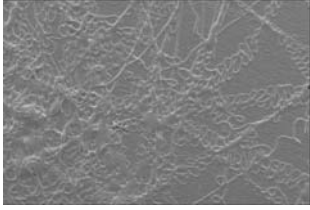
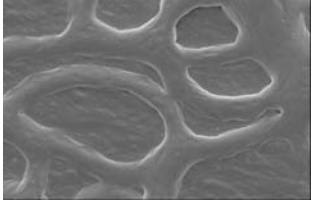
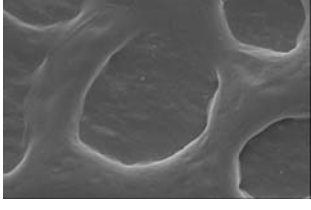
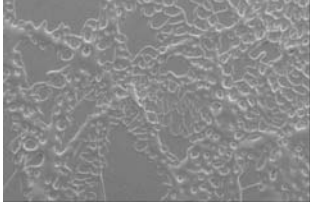
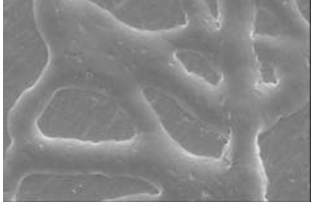
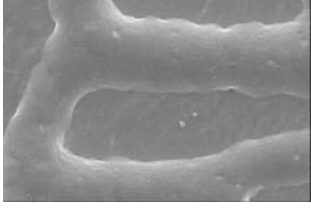
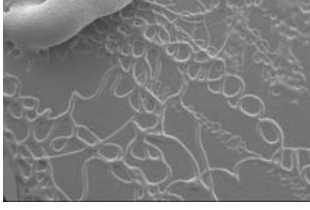
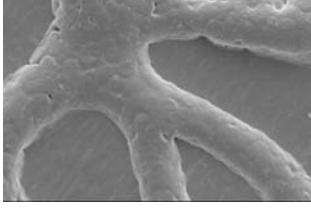
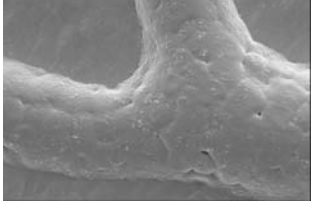


1			
2			
5			
7.5			

Taula F.8 Imatges de SEM per a l'estudi de l'efecte del cabal en el copolímer PEO₅₀₀₀-*b*-PCL₂₀₀₀₀

Cabal [ml/h]	50 x	500x	1000x
0.02			
0.05			
0.1			



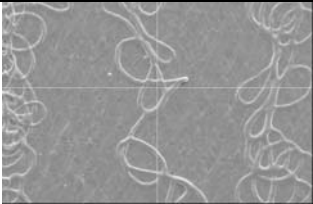
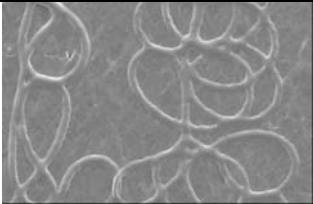
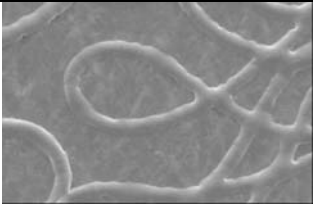
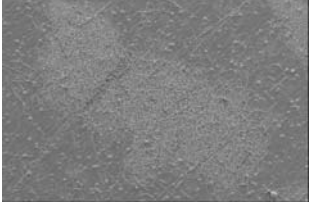
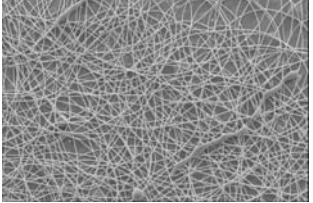
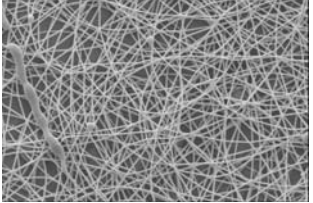
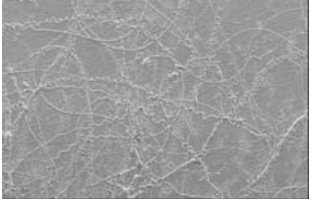

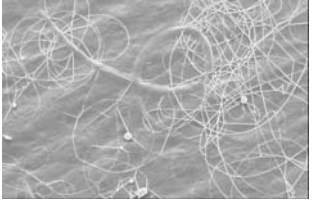
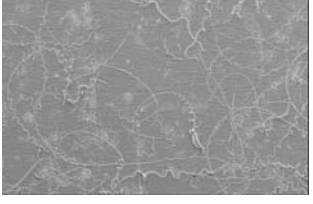
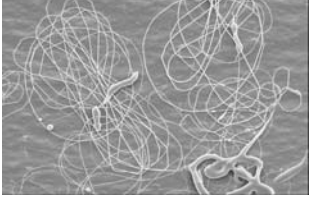
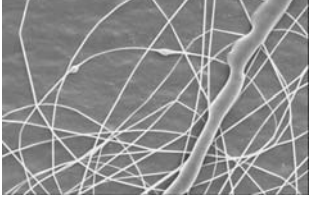
0.2			
0.5			
1			
2			
5			
7.5			



F.3 Estudi del paràmetre voltatge

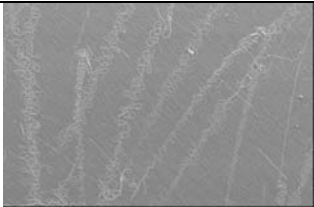
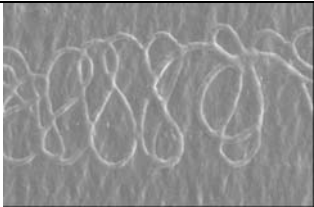
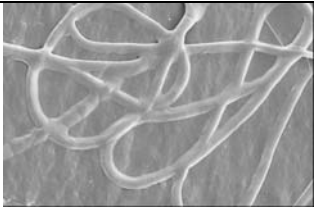
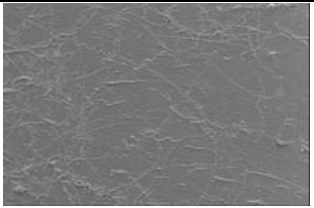
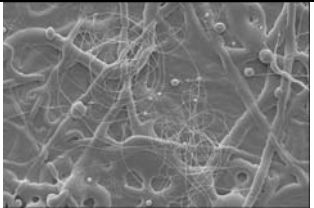
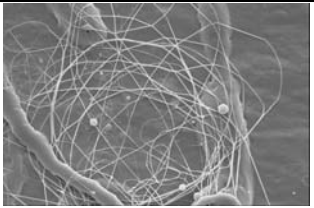
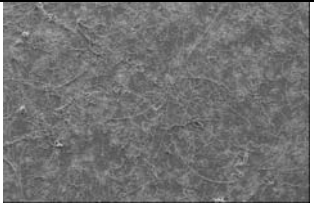
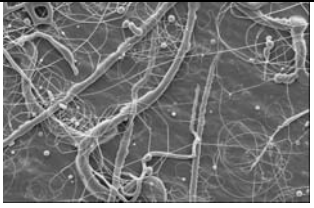
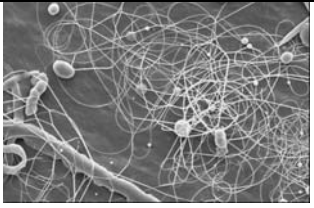
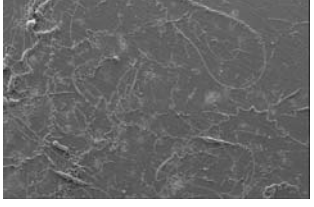
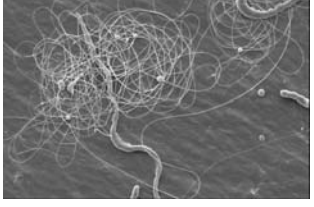
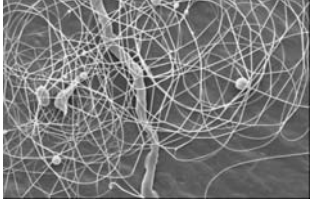
El paràmetre voltatge s'ha estudiat només entre 15 kV i 30 kV per als copolímers PEO₅₀₀₀-*b*-PCL₁₅₀₀₀ i PEO₅₀₀₀-*b*-PCL₂₀₀₀₀. S'han produït mostres a 0.05, 0.1 i 0.2 ml/h, els cabals que donen més fibres seques (partint dels resultats de paràmetre del cabal). El copolímer PEO₅₀₀₀-*b*-PCL₂₀₀₀₀ no dóna lloc a *electrospinning* a 15 kV. S'ha treballat a 85 °C.

Taula F.9 Imatges de SEM per a l'estudi de l'efecte del voltatge en el copolímer PEO₅₀₀₀-*b*-PCL₁₅₀₀₀ a 0.05 ml/h

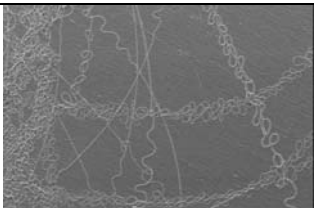
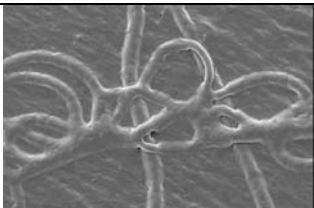
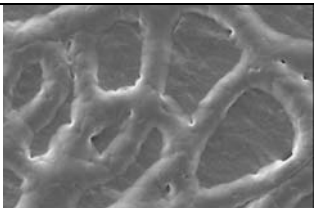
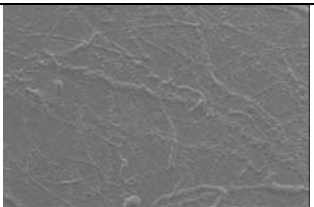
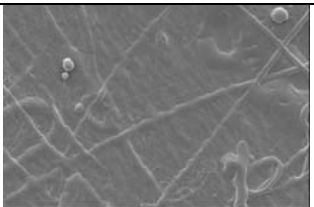
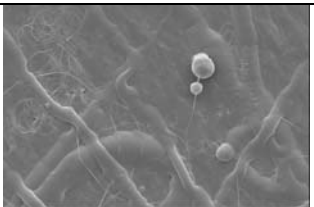
Voltatge [kV]	50 x	500x	1000x
15			
20			
25			
30			



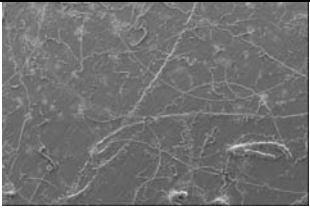
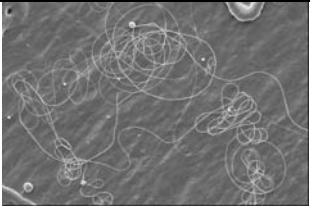
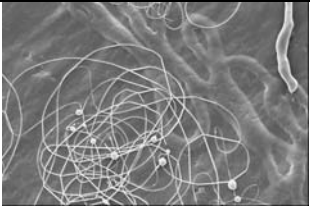
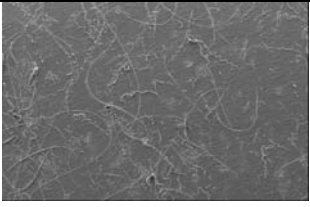

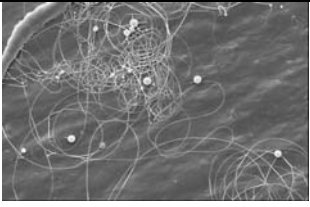
Taula F.10 Imatges de SEM per a l'estudi de l'efecte del voltatge en el copolímer PEO₅₀₀₀-*b*-PCL₁₅₀₀₀ a 0.1 ml/h

Voltatge [kV]	50 x	500x	1000x
15			
20			
25			
30			

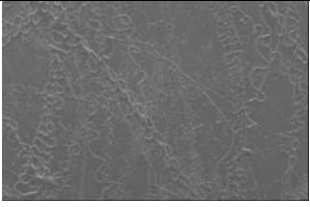
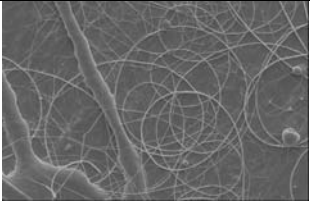
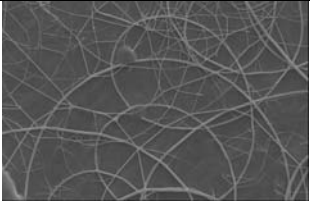
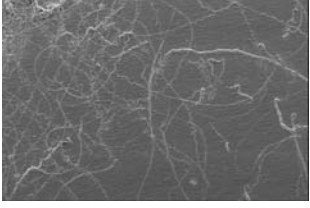
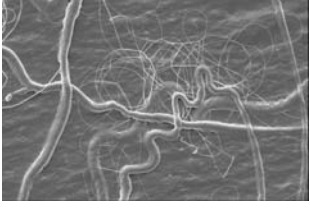

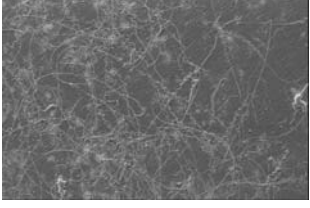
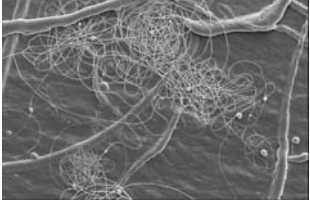

Taula F.11 Imatges de SEM per a l'estudi de l'efecte del voltatge en el copolímer PEO₅₀₀₀-*b*-PCL₁₅₀₀₀ a 0.2 ml/h

Voltatge [kV]	50 x	500x	1000x
15			
20			



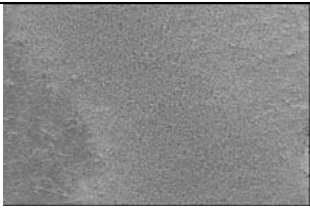
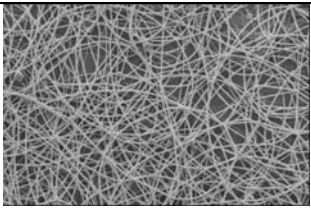
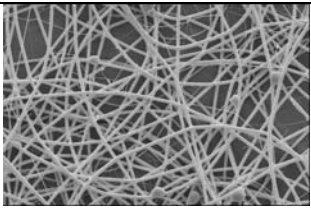
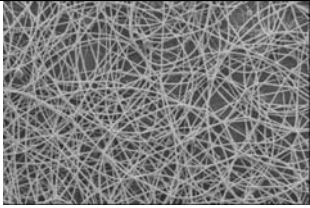
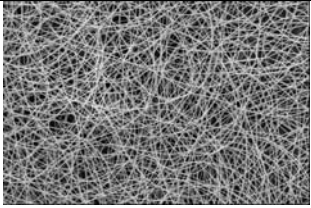
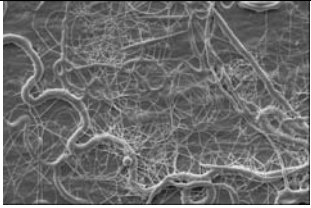
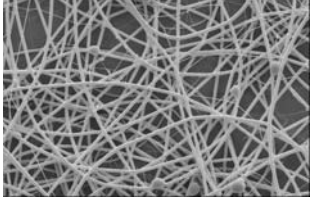
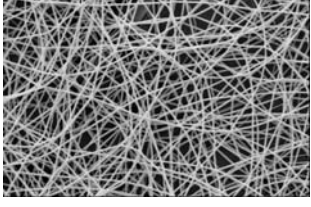
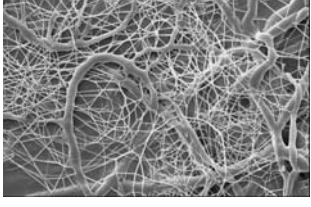
25			
30			

Taula F.12 Imatges de SEM per a l'estudi de l'efecte del voltatge en el copolímer PEO₅₀₀₀-*b*-PCL₂₀₀₀₀ a 0.05 ml/h

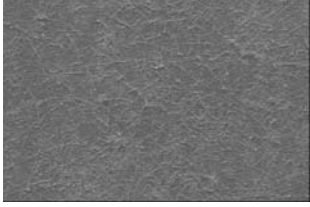

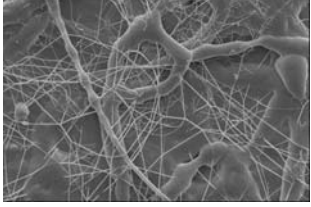
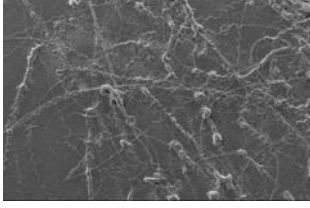
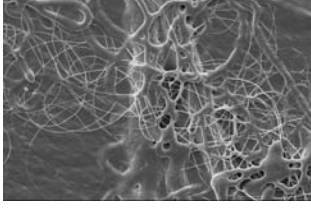
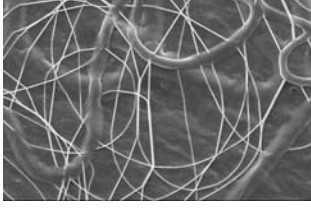
Voltatge [kV]	50 x	500x	1000x
20			
25			
30			



Taula F.13 Imatges de SEM per a l'estudi de l'efecte del voltatge en el copolímer PEO₅₀₀₀-*b*-PCL₂₀₀₀₀ a 0.1 ml/h

Voltatge [kV]	50 x	500x	1000x
20			
25			
30			

Taula F.14 Imatges de SEM per a l'estudi de l'efecte del voltatge en el copolímer PEO₅₀₀₀-*b*-PCL₂₀₀₀₀ a 0.2 ml/h

Voltatge [kV]	50 x	500x	1000x
20			
25			
30	